



# The Effect of Platelet-Rich Plasma on the Chondrogenic and Osteogenic Differentiation of Stem Cells

Andrea Pantalone<sup>1\*</sup>, Alessio Giannetti<sup>1</sup>, Daniele Vanni<sup>1</sup>, Sandra Verna<sup>2</sup>, Patrizia Di Gregorio<sup>2</sup> and Vincenzo Salini<sup>1</sup>

<sup>1</sup>Department of Medicine and Science of Aging, University of Study "G. d'Annunzio", Italy

<sup>2</sup>Department of Immunohaematology and Transfusional Medicine Service, "Ss. Annunziata" Hospital, Italy

## Abstract

Tissue engineering is intended to regenerate the damaged organs or to promote their healing. The techniques used by this branch of regenerative medicine consist in biological active molecules, stem cells and scaffolds. Platelet-Rich Plasma (PRP) is a plasma fraction containing an amount of platelets several times above the baseline. In the last decades the use of PRP has gained popularity in the field of the tissue engineering. Its properties are anti-inflammatory, analgesic and neo-angiogenic; for this reason it is able to promote tissue formation and remodeling by influencing stem cells migration, proliferation and differentiation. Stem cells used in tissue engineering are harvested from different sources. In orthobiologics, the application of the regenerative medicine in the orthopedic field, Mesenchymal Stem Cells (MSCs) from bone marrow are the most used. Nevertheless there are other alternative sources of MSCs that can be chosen such as adipose tissue, synovial fluid, muscle tissue, umbilical cord and amniotic fluid. The aim of this mini-review is to collect the works present in literature where PRP has been used in combination with stem cells from different origins: this association has shown promising results both *in vitro*, in chondrogenic and osteogenic differentiation and *in vivo*, in osteochondral defects healing.

**Keywords:** Platelet rich plasma; Orthobiologics; Stem cells; Mesenchymal stem cells

## OPEN ACCESS

### \*Correspondence:

Andrea Pantalone, Department of Medicine and Science of Aging, University of Study "G. d'Annunzio" Chieti-Pescara, "Ss. Annunziata" Hospital, Italy, Via dei Vestini 35, 66013 Chieti, Tel: (39) 0871 358263; Fax: (39) 0871 560082; E-mail: pantaloneandrea@libero.it

Received Date: 20 Nov 2017

Accepted Date: 12 Jan 2018

Published Date: 19 Jan 2018

### Citation:

Pantalone A, Giannetti A, Vanni D, Verna S, Di Gregorio P, Salini V. The Effect of Platelet-Rich Plasma on the Chondrogenic and Osteogenic Differentiation of Stem Cells. *Annals Stem Cell Regenerat Med.* 2018; 1(1): 1003.

Copyright © 2018 Andrea Pantalone.

This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

## Introduction

### Platelet-Rich Plasma (PRP)

Platelet-Rich Plasma (PRP) consists in a plasma fraction containing an amount of platelets several times more than the normal platelet count (150 000-350 000/ $\mu$ l) [1-4]. Platelets or thrombocytes are biconvex discoid elements of the blood (2-3 $\mu$ m diameter), represented by fragments of cytoplasm originating from megakaryocytes in the bone marrow [1,2,5]. About 40 years ago, platelets were only known as hemostatic cells until, in 1974, Ross et al. [6] noticed that the addition of activated platelets into a smooth muscle cell culture increased the mitogenic activity of the cells. Thrombocytes lack the nuclei and some organelles such as the Golgi apparatus and the endoplasmic reticulum, but they have structures like  $\alpha$ ,  $\delta$ ,  $\lambda$  granules [7]. Granules  $\alpha$  contain more than 30 bioactive proteins such as PDGF, TGF- $\beta$ , PF-4, IL-1, PDAF, VEGF, EGF, PDEGF, ECGF, IGF, osteocalcin, osteonectin, fibrinogen, vitronectin, fibronectin, and thrombospondin [1,2,4,8-18]. These factors are released after platelet activation, 70% of them within 10 minutes and after 1 hour the 95% of the proteins are released [2,5,18-20].

The interest in the use of PRP depends on the presence of these cytokines, chemokines and growth factors. The PRP activation is usually obtained by adding calcium chloride, autologous or bovine thrombin, batroxobin and collagen type I [1,2,9,18,21-23]. For the *in vitro* use, the activation of PRP is imperative, on the other hand for the *in vivo* use, the activation is obtained in situ by the effect of the endogenous collagen and thromboplastin of the connective tissue [2,3,8,18, 20, 21,24-27]. PRP is produced by centrifugating the whole blood in order to separate the various components according to their own density. There are different techniques to prepare the PRP [8,28] and there are different kinds of PRP as well, such as Pure PRP or leukocytes-poor platelet-rich plasma, and leukocytes-rich platelet-rich plasma [29-31]. This latter seems to have some antibacterial properties but, at same time, the leukocytes in PRP releases pro-inflammatory cytokines like IL-1 $\beta$  and TNF- $\alpha$  that, activating the NF- $\kappa$ B pathway, inhibit tissue healing [8,20,32-38]. In the last decades the use of PRP has gained popularity in the field of the tissue engineering because of its anti-inflammatory and analgesic properties, its capacity to induce neo-angiogenesis, tissue formation and remodeling

by influencing stem cells migration, proliferation and differentiation [2,8,9,21,28,32,39-48].

PRP is being used in different applications in medicine such as in the area of aesthetic medicine, maxillofacial surgery and orthopedics [8,49-52]. Nevertheless, the evidences in the literature of the use of PRP are conflicting, in fact in some studies the use of PRP leads to benefits while in others the effect of PRP is not relevant [8]. On the other hand the combination of PRP and stem cells, harvested from different sources, represents a new promising branch of the orthobiology giving encouraging results both *in vitro* and *in vivo*.

### Stem cells

Adult stem cells have a central role in regenerative medicine, they in fact dominate the market compared to the embryonic stem cells because of their ease of isolation, that does not give rise to ethical controversies, because of their multipotentiality and their capacity of self-renewal, lower maintenance cost, lower risk of induction of teratomas after *in vivo* transplantation and better immunocompatibility [53-62].

Mesenchymal Stem Cells (MSC) could be harvested from different sources such as bone marrow, adipose tissue, peripheral blood, lungs, synovial membrane, dental pulp, satellite muscle cells, placenta, umbilical cord and cord blood [53,63-77]. Furthermore it has to be said that the properties, markers expression, differentiation capacities, efficiency and paracrine functions change according to the source of stem cells chosen. Therefore the choice of the type of stem cells could depend on the clinical application [53,78-86].

### PRP and Bone Marrow Stem Cells (BMSCs)

BMSCs are the most studied and used cells in regenerative medicine [53]. These cells are isolated from the heterogeneous cell population present in the bone marrow aspiration. The isolation of the stem cells is obtained using a culture protocol in which the non-adherent cells are eliminated with the medium changes and after 9-10 days most of the cells remained are mesenchymal stem cells [87]. There are different methods of isolation with different degrees of sophistication [87-91].

In the definition of MSC there is the capacity to differentiate into mesodermal lineages like fat, bone and cartilage. However when MSCs are co-cultured with specific inducing factors, they demonstrated to have endodermic and neuro-ectodermic differentiation potential as well [87,92-96].

The combination of PRP and BMSCs could be an alternative to the autologous or allogenic graft for osteochondral lesions [97,98]. This association has often been used in orthobiologics to face the most common diseases in orthopedics such as cartilage and bone defects [99-102].

A recent study from Wei et al. [103] has demonstrated that using PRP and BMSCs for the treatment of osteoporosis in ovariectomized rats has led to better results than using PRP or BMSCs alone. Therefore this study suggests an alternative strategy to treat osteoporotic bone loss and heal bone defects in general.

PRP and BMSCs have been used together also with a combination of scaffolds, as well as done by Li et al. [104], that successfully managed osteochondral defects in beagles.

### PRP and Adipose-derived Stem Cells (ADSCs)

ADSCs could represent a good alternative to the BMSCs for their ease of isolation, abundance, proliferation and differentiation properties into different cell lineages (adipogenic, chondrogenic, osteogenic, myogenic, neurogenic lineage) [105,106-110]. ADSCs can be obtained by lipoaspirates from patients inguinal fat pads. Stem cells from adipose tissue are more numerous than those from other stem cells sources (10<sup>7</sup> ADSCs from 300 ml of lipoaspirate) [105,111,112].

The growth factors founded in the PRP can influence the migration, proliferation and differentiation of ADSCs. In addition the fibrin network in the PRP works as a scaffold. Thereby this cooperation can certainly be used in the regenerative medicine [105,113-119].

In orthopedics PRP and ADSCs have been largely investigated both for bone defects and cartilage lesions [120-123]. Tajima et al. [124] demonstrated that bone healing in rats was significantly greater when PRP and ADSCs were used together instead of alone. In a similar way Van Pham et al. [125] showed with their work the capacity of ADSCs and PRP of promoting injured particular cartilage healing that was more efficient than the one with untreated ADSCs.

### PRP and Synovial Fluid Mesenchymal Stem Cells (SF-MSCs)

Stem cells from human synovial membrane were isolated for the first time in 2001 by De Bari et al. [126]. Likely all the MSCs SF-MSCs have high proliferation capacities and they are able to differentiate into adipogenic, chondrogenic and osteogenic lineages [127,128] however SF-MSCs show greater chondrogenesis compared to the stem cells from other tissues [127,129]. Furthermore SF-MSCs isolation is easier and less invasive than other MSCs; in fact it is obtained by a simple Arthrocentesis. The number of stem cells that can be found in the synovial fluid is greater if the patient is affected by osteoarthritis, anterior cruciate ligament injuries or meniscus lesions [127,130-133].

The effectiveness of the combination of PRP and SF-MSCs has been proved both *in vitro* and *in vivo*. *In vitro* the chondrogenic and osteogenic differentiation was obtained [127], while the *in vivo* study performed in an animal model, has shown good results on the treatment of damaged articular cartilage in rabbits [133].

### PRP and Muscle-Derived Stem Cells (MDSCs)

MDSCs or Muscle Satellite Cells were described for the first time by Mauro [134,135]. Their capacity of self-renewal, long-term proliferation, and multi-lineage differentiation potential both *in vitro* and *in vivo* has been demonstrated [135,136-138].

Although the literature is poor of studies concerning the combination of PRP and MDSCs, some authors like [139] and Huang et al. [135] have investigated this association and they affirm the both *in vitro* and *in vivo* efficacy of PRP in promoting proliferation, osteogenic and chondrogenic differentiation in Muscle Satellite Cells.

### PRP and Umbilical Cord Stem Cells (UCSCs)

Mesenchymal Stem Cells obtained from the umbilical cord blood are already known for their self-renewal capacity and their ability to differentiate into adipose, muscle, bone and cartilage tissue [140-145].

Theoretically because of their immunologic immaturity, UCSCs can offer some advantages compared with the stem cells from bone marrow aspirates, but on the other hand, the low quantity of stem cells present in a single sample from the umbilical cord blood could represent a limitation for their clinical application in an adult population with high body weight [146]. Despite this observation [140] and [147] have believed in the potential of the use of UCSCs in the regenerative medicine and demonstrated their ability to differentiate into bone lineage *in vitro* and to accelerate bone regeneration *in vivo* when combined with PRP.

### PRP and Amniotic Fluid Stem Cells (AFSCs)

Amniotic fluid could represent a new source from where harvest stem cells. AFSCs are collected through amniocentesis in women undergoing prenatal diagnosis (16<sup>th</sup>-18<sup>th</sup> week of pregnancy) [98,148-155]. These cells possess interesting characteristics; in fact AFSCs have intermediate properties between embryonic stem cells and adult stem cells. Indeed AFSCs present a greater potency than adult stem cells while, comparing embryonic fluid stem cells with AFSCs, these latter are easier to collect and more ethically accept because no embryo needs to be suppressed. In addition they are more genetically stable and therefore they do not induce teratomas after *in vivo* transplantation [98,156-159].

For the use in orthobiologics AFSCs may represent a new alternative, considering their ability to differentiate *in vitro* into osteogenic precursors, in fact it was demonstrated that 85% of AFSCs versus 50% of MSCs lead to the formation of osteogenic colonies [98,159-167]. Wang et al. [168] have investigated the AFSCs *in vivo* potential as well; in fact in their work they obtained a promotion of maxillary alveolar bone defect in rats combining AFSCs and PRP.

### References

- Fernandes G, Yang S. Application of platelet-rich plasma with stem cells in bone and periodontal tissue engineering. *Bone Res.* 2016;4:16036.
- Alousou J, Thompson M, Hulley P, Noble A, Willett K. The biology of platelet-rich plasma and its application in trauma and orthopaedic surgery. *J Bone Joint Surg [Br].* 2009;91(8):987-96.
- Dhillon MS, Behera P, Patel S, Shetty V. Orthobiologics and platelet rich plasma. *Indian J Orthop.* 2014;48(1):1-9.
- Marx RE. Platelet-rich plasma (PRP): what is PRP and what is not PRP? *Implant Dent.* 2001;10(4):225-8.
- Harrison P. Platelet function analysis. *Blood Reviews.* 2005;19(2):111-23.
- Ross R, Glomset J, Kariya B, Harker L. A platelet-dependent serum factor that stimulates the proliferation of arterial smooth muscle cells *in vitro*. *Proc Natl Acad Sci USA.* 1974;71(4):1207-10.
- Harrison P, Cramer EM. Platelet alpha-granules. *Blood Rev.* 1993;7(1):52-62.
- Hussain N, Johal H, Bhandari M. An evidence-based evaluation on the use of platelet rich plasma in orthopedics - a review of the literature. *Sicot J.* 2017;3:57.
- Zahn J, Loibl M, Sprecher C, Nerlich M, Alimi M, Verrier S, et al. Platelet-Rich Plasma as an Autologous and Proangiogenic Cell Delivery System. *Mediators Inflamm.* 2017;2017:1075975.
- Senzel L, Gnatenko DV, Bahou WF. The platelet proteome. *Curr Opin Hematol.* 2009;16(5):329-33.
- Zahedi RP, Lewandrowski U, Wiesner J, Wortelkamp S, Moebius J, Schütz C, et al. *J Proteome Res.* 200;7(2):526-34.
- Ferrari M, Zia S, Valbonesi M, Henriquet F, Venere G, Spagnolo S, et al. A new technique for hemodilution, preparation of autologous platelet-rich plasma and intraoperative blood salvage in cardiac-surgery. *Int J Artif Organs.* 1987;10(1):47-50.
- Sanchez AR, Sheridan PJ, Kupp LI. Is platelet-rich plasma the perfect enhancement factor: a current review. *Int J Oral Maxillofac Implants.* 2003;18(1):93-103.
- Marx RE, Carlson ER, Eichstaedt RM, Schimmele SR, Strauss JE, Georgeff KR. Platelet rich plasma: growth factor enhancement for bone grafts. *Oral Surg Oral Med Oral Pathol Oral RadiolEndod.* 1998;85(6):638-46.
- Bucholz RW, Einhorn TA, Marsh JL. Bone and joint healing. In: Bucholz RW, Heckman JD, Court-Brown C, eds. *Rockwood & Green's fractures in adults.* Sixth ed. Lippincott Williams & Wilkins. 2006. p. 300-11.
- King SM, Reed GL. Development of platelet secretory granules. *Semin Cell Dev Biol.* 2002;13(4):293-302.
- Wasterlain AS, Braun HJ, Harris AH, Kim HJ, Dragoo JL. The systemic effects of platelet-rich plasma injection. *Am J Sports Med.* 2013;41(1):186-93.
- Marx RE. Platelet-rich plasma: evidence to support its use. *J Oral Maxillofac Surg.* 2004;62(4):489-96.
- Arnoczky SP, Delos D, Rodeo SA. What is Platelet-Rich plasma? *Operative Techniques in Sports Med.* 2011;19(3):142-8.
- Arnoczky SP, Sheibani-Rad S. The basic science of platelet-rich plasma (PRP): what clinicians need to know. *Sports Med Arthrosc Rev.* 2013;21(4):180-5.
- Marmotti A, Rossi R, Castoldi F, Roveda E, Michielon G, Peretti GM. PRP and Articular Cartilage: A Clinical Update. *Biomed Res Int.* 2015;2015:542502.
- Evans CH. Advances in regenerative orthopedics. *Mayo Clin Proc.* 2013;88(11):1323-39.
- Martinez CE, Gonzalez SA, Palma V, Smith PC. Platelet-poor and platelet-rich plasma stimulate bone lineage differentiation in periodontal ligament stem cells. *J Periodontol.* 2016;87(2):e18-26.
- Hsu WK, Mishra A, Rodeo SR, Fu F, Terry MA, Randelli P, et al. Platelet-rich plasma in orthopaedic applications: evidence-based recommendations for treatment. *J Am Acad Orthop Surg.* 2013;21(12):739-48.
- Busilacchi A, Gigante A, Mattioli-Belmonte M, Manzotti S, Muzzarelli RAA. Chitosan stabilizes platelet growth factors and modulates stem cell differentiation toward tissue regeneration. *Carbohydr Polym.* 2013;98(1):665-76.
- Kutlu B, Aydın RST, Akman AC, Gümüşderelioglu M, Nohutcu RM. Platelet-rich plasma-loaded chitosan scaffolds: preparation and growth factor release kinetics. *J Biomed Mater Res B Appl Biomater.* 2013;101(1):28-35.
- Mishra A, Harmon K, Woodall J, Vieira A. Sports medicine applications of platelet rich plasma. *Curr Pharm Biotechnol.* 2012;13(7):1185-95.
- Eppley BL, Pietrzak WS, Blanton M. Platelet-rich plasma: a review of biology and applications in plastic surgery. *Plast Reconstr Surg.* 2006;118(6):147-159.
- Dohan Ehrenfest DM, Andia I, Zumstein MA, Zhang CQ, Pinto NR, Bielecki T. Classification of platelet concentrates (Platelet-Rich Plasma-PRP, Platelet-Rich Fibrin- PRF) for topical and infiltrative use in orthopedic and sports medicine: current consensus, clinical implications and perspectives. *Muscle Ligaments Tendons J.* 2014;4(1):3-9.
- Dohan Ehrenfest DM, Bielecki T, Del Corso M, Inchingolo F, Sammartino G. Shedding light in the controversial terminology for platelet-rich products: platelet-rich plasma (PRP), platelet-rich fibrin (PRF), platelet-leukocyte gel (PLG), preparation rich in growth factors

- (PRGF), classification and commercialism. *J Biomed Mater Res A*. 2010;95(4):1280-2.
31. Dohan Ehrenfest DM, Bielecki T, Jimbo R, Barbe G, Del Corso M, Inchingolo F, Sammartino G. Do the fibrin architecture and leukocyte content influence the growth factor release of platelet concentrates? An evidence-based answer comparing a pure platelet-rich plasma (P-PRP) gel and a leukocyte- and platelet-rich fibrin (L-PRF). *Curr Pharm Biotechnol*. 2012;13(7):1145-52.
  32. Schär MO, Diaz-Romero J, Kohl S, Zumstein MA, Nestic D. Platelet-rich concentrates differentially release growth factors and induce cell migration *in vitro*. *Clin Orthop Relat Res*. 2015;473(5):1635-43.
  33. Filardo G, Kon E, Pereira Ruiz MT, Vaccaro F, Guitaldi R, Di Martino A, et al. Platelet-rich plasma intra-articular injections for cartilage degeneration and osteoarthritis: single- versus doublespinning approach. *Knee Surg Sports Traumatol Arthrosc*. 2012;20(10):2082-91.
  34. Riboh JC, Saltzman BM, Yanke AB, Fortier L, Cole BJ. Effect of Leukocyte Concentration on the Efficacy of Platelet-Rich Plasma in the Treatment of Knee Osteoarthritis. *Am J Sports Med*. 2016;44:792-800.
  35. McCarrel TM, Minas T, Fortier LA. Optimization of leukocyte concentration in platelet-rich plasma for the treatment of tendinopathy. *J Bone Joint Surg Am*. 2012;94(19):e143(1-8).
  36. Singh R, Ahmed S, Islam N, Goldberg VM, Haqqi TM. Epigallocatechin-3-gallate inhibits interleukin-1 $\beta$ -induced expression of nitric oxide synthase and production of nitric oxide in human chondrocytes: suppression of nuclear factor kappa B activation by degradation of the inhibitor of nuclear factor kappa B. *Arthritis Rheum*. 2002;46(8):2079-86.
  37. Liacini A, Sylvester J, Li WQ, Zafarullah M. Inhibition of interleukin-1-stimulated MAP kinases, activating protein-1 (AP-1) and nuclear factor kappa B (NF-kappa B) transcription factors down-regulates matrix metalloproteinase gene expression in articular chondrocytes. *Matrix Biol*. 2002;21(3):251-62.
  38. Ishinaga H, Jono H, Lim JH, Komatsu K, Xu X, Lee J, et al. Synergistic induction of nuclear factor-kB by transforming growth factor-beta and tumour necrosis factor-alpha is mediated by protein kinase A-dependent RelA acetylation. *Biochem J*. 2009;417(2):583-91.
  39. Bendinelli P, Matteucci E, Dogliotti G, Corsi MM, Banfi G, Maroni P, et al. Molecular basis of anti-inflammatory action of platelet-rich plasma on human chondrocytes: mechanisms of NF-B inhibition via HGF. *J Cell Physiol*. 2010;225(3):757-66.
  40. Krüger JP, Hondke S, Endres M, Pruss A, Siclari A, Kaps C. Human platelet-rich plasma stimulates migration and chondrogenic differentiation of human subchondral progenitor cells. *J Orthop Res*. 2012;30(6):845-52.
  41. Rubio-Azpeitia E, Andia I. Partnership between platelet-rich plasma and mesenchymal stem cells: *in vitro* experience. *Muscles Ligaments and Tendons J*. 2014;4(1):52-62.
  42. Vogl M, Fischer J, Jäger M, Zilkens C, Krauspe R, Herten M. Can thrombin-activated platelet releasate compensate the age-induced decrease in cell proliferation of MSC? *J Orthop Res*. 2013;31(11):1786-95.
  43. Lippross S, Moeller B, Haas H, Tohidnezhad M, Steubesand N, Wruck CJ, et al. Intraarticular injection of platelet-rich plasma reduces inflammation in a pig model of rheumatoid arthritis of the knee joint. *Arthritis Rheum*. 2011;63(11):3344-53.
  44. Liu J, Song W, Yuan T, Xu Z, Jia W, Zhang C. A comparison between platelet-rich plasma (PRP) and hyaluronate acid on the healing of cartilage defects. *PLoS ONE*. 2014;9(5):e97293.
  45. Filardo G, Kon E, Roffi A, Di Matteo B, Merli ML, Marcacci M. Platelet-rich plasma: why intra-articular? A systematic review of preclinical studies and clinical evidence on PRP for joint degeneration. *Knee Surg Sports Traumatol Arthrosc*. 2015;23(9):2459-74.
  46. Boswell SG, Cole BJ, Sundman EA, Karas V, Fortier LA. Platelet-rich plasma: a milieu of bioactive factors. *Arthroscopy*. 2012;28(3):429-39.
  47. Thushara RM, Hemshekhar M, Basappa, Kemparaju K, Rangappa KS, Girish KS. Biologicals, platelet apoptosis and human diseases: an outlook. *Crit Rev Oncol Hematol*. 2015;93(3):149-58.
  48. Schliephake H. Bone growth factors in maxillofacial skeletal reconstruction. *Int J Oral Maxillofac Surg*. 2002;31(5):469-84.
  49. Sadick NS. New-Generation Therapies for the Treatment of Hair Loss in Men. *Dermatol Clin*. 2017;36(1):63-7.
  50. Ghoddusi J, Maghsudlu A, Jafarzadeh H, Jafarian A, Forghani M. Histological Evaluation of the Effect of Platelet-rich Plasma on Pulp Regeneration in Nonvital Open Apex Teeth: An Animal Study. *J Contemp Dent Pract*. 2017;18(11):1045-50.
  51. Parra F, Morales-Rome DE, Campos-Rodríguez R, Cruz-Hernández TR, Drago-Serrano ME. Effect of platelet-rich plasma on patients after blepharoplasty surgery. *Orbit*. 2017;12:1-6.
  52. Bousnaki M, Bakopoulou A, Koidis P. Platelet-rich plasma for the therapeutic management of temporomandibular joint disorders: a systematic review. *Int J Oral Maxillofac Surg*. 2018;47(2):188-198.
  53. Samsonraj RM, Raghunath M, Nurcombe V, Hui JH, van Wijnen AJ, Cool SM. Concise Review: Multifaceted Characterization of Human Mesenchymal Stem Cells for Use in Regenerative Medicine. *Stem Cells Transl Med*. 2017;6(12):2173-85.
  54. Caplan AI. Mesenchymal stem cells. *J Orthop Res*. 1991;9(5):641-50.
  55. Corselli M, Chin CJ, Parekh C, Sahaghian A, Wang W, Ge S, et al. Perivascular support of human hematopoietic stem/progenitor cells. *Blood*. 2013;121(15):2891-901.
  56. Crisan M, Yap S, Casteilla L, Chen CW, Corselli M, Park TS, et al. A perivascular origin for mesenchymal stem cells in multiple human organs. *Cell Stem Cell*. 2008;3(3):301-13.
  57. Blocki A, Wang Y, Koch M, Peh P, Beyer S, Law P, et al. Not all MSCs can act as pericytes: Functional *in vitro* assays to distinguish pericytes from other mesenchymal stem cells in angiogenesis. *Stem Cells Dev*. 2013;22(17):2347-55.
  58. Sacchetti B, Funari A, Michienzi S, Di Cesare S, Piersanti S, Saggio I, et al. Self-renewing osteoprogenitors in bone marrow sinusoids can organize a hematopoietic microenvironment. *Cell*. 2007;131(2):324-36.
  59. Eaker S, Armant M, Brandwein H, Burger S, Campbell A, Carpenito C, et al. Concise review: Guidance in developing commercializable autologous/patient-specific cell therapy manufacturing. *Stem Cells Transl Med*. 2013;2(11):871-83.
  60. Muller-Cohn J, Diaz P, Muller R. Stem cell value chains. In: Verte` s AA, Qureshi N, Caplan AI et al. e. *Stem Cells in Regenerative Medicine*. Chichester, UK: John Wiley. 2015;p:341-54.
  61. Schnitzler AC, Verma A, Kehoe DE, Jing D, Murrel JR, Der KA, et al. Bioprocessing of human mesenchymal stem/stromal cells for therapeutic use: Current technologies and challenges. *Biochem Eng J*. 2016;108:3-13.
  62. Research GV. Stem cells market size, global industry research report, 2014-2025. Report ID: 978-1-68038-130-6.
  63. Aust L, Devlin B, Foster SJ, Halvorsen YD, Hicok K, du Laney T, et al. Yield of human adipose-derived adult stem cells from liposuction aspirates. *Cytotherapy*. 2004;6(1):7-14.
  64. Smiler D, Soltan M, Albitar M. Toward the identification of mesenchymal stem cells in bone marrow and peripheral blood for bone regeneration. *Implant Dent*. 2008;17(3): 236-47.
  65. He Q, Wan C, Li G. Concise review: Multipotent mesenchymal stromal cells in blood. *Stem Cells*. 2007;25(1):69-77.
  66. Griffiths MJD, Bonnet D, Janes SM. Stem cells of the alveolar epithelium. *Lancet*. 2005;366(9481):249-60.

67. Tuli R, Li WJ, Tuan RS. Current state of cartilage tissue engineering. *Arthritis Res Ther.* 2003;5(5):235-38.
68. Fan J, Varshney RR, Ren L, Cai D, Wang DA. Synovium- derived mesenchymal stem cells: A new cell source for musculoskeletal regeneration. *Tissue Eng Part B Rev.* 2009;15(1):75-86.
69. Gay IC, Chen S, MacDougall M. Isolation and characterization of multipotent human periodontal ligament stem cells. *Orthod Craniofac Res.* 2007;10(3):149-60.
70. Jackson WM, Nesti LJ, Tuan RS. Potential therapeutic applications of muscle-derived mesenchymal stem and progenitor cells. *Expert Opin Biol Ther.* 2010;10(4):505-17.
71. In't Anker PS, Scherjon SA, Kleijburg-van der Keur C, De Groot-Swings GM, Claas FH, Fibbe WE, et al. Isolation of mesenchymal stem cells of fetal or maternal origin from human placenta. *Stem Cells.* 2004;22(7):1338-45.
72. Miao Z, Jin J, Chen L, Zhu J, Huang W, Zhao J, et al. Isolation of mesenchymal stem cells from human placenta: Comparison with human bone marrow mesenchymal stem cells. *Cell Biol Int.* 2006;30:681-7.
73. Corrao S, La Rocca G, Lo Iacono M, Corsello T, Farina F, Anzalone R. Umbilical cord revisited: From Wharton's jelly myofibroblasts to mesenchymal stem cells. *Histol Histopathol.* 2013;28(10):1235-44.
74. Erices A, Conget P, Minguell JJ. Mesenchymal progenitor cells in human umbilical cord blood. *Br J Haematol.* 2000;109(1):235-42.
75. Mareschi K, Biasin E, Piacibello W, Aglietta M, Madon E, Fagioli F. Isolation of human mesenchymal stem cells: Bone marrow versus umbilical cord blood. *Haematologica.* 2001;86(10):1099-100.
76. Seo BM, Miura M, Gronthos S, Bartold PM, Batouli S, Brahimi J, et al. Investigation of multipotent postnatal stem cells from human periodontal ligament. *Lancet.* 2004;364(9429):149-55.
77. Shi S, Gronthos S. Perivascular niche of postnatal mesenchymal stem cells in human bone marrow and dental pulp. *J Bone Miner Res.* 2003;18(4):696-704.
78. Elahi KC, Klein G, Avci-Adali M, Sievert KD, MacNeil S, Aicher WK. Human Mesenchymal Stromal Cells from Different Sources Diverge in Their Expression of Cell Surface Proteins and Display Distinct Differentiation Patterns. *Stem Cells Int.* 2016;2016:5646384.
79. Kwon A, Kim Y, Kim M, Kim J, Choi H, Jekarl DW, et al. Tissue-specific differentiation potency of mesenchymal stromal cells from perinatal tissues. *Sci Rep.* 2016;6:23544.
80. Davies JE, Walker JT, Keating A. Concise review: Wharton's Jelly: The rich, but enigmatic, source of mesenchymal stromal cells. *Stem Cells Transl Med.* 2017;6(7):1620-30.
81. Chen JY, Mou XZ, Du XC, Xiang C. Comparative analysis of biological characteristics of adult mesenchymal stem cells with different tissue origins. *Asian Pac J Trop Med.* 2015;8(9):739-46.
82. Billing AM, Ben Hamidane H, Dib SS, Cotton RJ, Bhagwat AM, Kumar P, et al. Comprehensive transcriptomic and proteomic characterization of human mesenchymal stem cells reveals source specific cellular markers. *Sci Rep.* 2016;6:21507.
83. Sakaguchi Y, Sekiya I, Yagishita K, Muneta T. Comparison of human stem cells derived from various mesenchymal tissues: Superiority of synovium as a cell source. *Arthritis Rheum.* 2005;52(8):2521-29.
84. Rider DA, Nalathamby T, Nurcombe V, Cool SM. Selection using the alpha-1 integrin (CD49a) enhances the multipotentiality of the mesenchymal stem cell population from heterogeneous one marrow stromal cells. *J Mol Histol.* 2007;38(5):449-58.
85. Hass R, Kasper C, Böhm S, Jacobs R. Different populations and sources of human mesenchymal stem cells (MSC): A comparison of adult and neonatal tissue-derived MSC. *Cell Commun Signal.* 2011;9:12.
86. Maleki M, Ghanbarvand F, Reza Behvarz M, Ejtemaei M, Ghadirkhomi E. Comparison of mesenchymal stem cell markers in multiple human adult stem cells. *Int J Stem Cells.* 2014;7(2):118-26.
87. Liu Z, Zhu Y, Ge R, Zhu J, He X, Yuan X. Combination of bone marrow mesenchymal stem cells sheet and platelet rich plasma for posterolateral lumbar fusion. *Oncotarget.* 2017;8(37):62298-311.
88. Baghaei K, Hashemi SM, Tokhanbigli S, Asadi Rad A, Assadzadeh-Aghdai H, Sharifian A, et al. Isolation, differentiation, and characterization of mesenchymal stem cells from human bone marrow. *Gastroenterol Hepatol Bed Bench.* 2017;10(3):208-13.
89. Grisendi G, Annerén C, Cafarelli L, Sternieri R, Veronesi E, Cervo GL, et al. GMP-manufactured density gradient media for optimized mesenchymal stromal/stem cell isolation and expansion. *Cytotherapy.* 2010;12(4):466-77.
90. Gronthos S, Zannettino AC, Hay S, Shi S, Graves SE, Kortessidis A, et al. Molecular and cellular characterisation of highly purified stromal stem cells derived from human bone marrow. *J Cell Sci.* 2003;116(Pt 9):1827-1835.
91. Miltenyi S, Muller W, Weichel W, Radbruch A. High gradient magnetic cell separation with MACS. *Cytometry.* 1990;11(2):231-8.
92. Sarugaser R, Hanoun L, Keating A, Tanford WL, Davies JE. Human mesenchymal stem cells self-renew and differentiate according to a deterministic hierarchy. *PLoS One.* 2009;4(8):e6498.
93. Kopen GC, Prockop DJ, Phinney DG. Marrow stromal cells migrate throughout forebrain and cerebellum, and they differentiate into astrocytes after injection into neonatal mouse brains. *Proc Natl Acad Sci USA.* 1999;96(19):10711-6.
94. Reger RL, Tucker AH, Wolfe MR. Differentiation and characterization of human MSCs. *Methods Mol Biol.* 2008;449:93-107.
95. Petersen BE, Bowen WC, Patrene KD, Mars WM, Sullivan AK, Murase N, et al. Bone marrow as a potential source of hepatic oval cells. *Science.* 1999;284(5417):1168-70.
96. Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, et al. Multilineage potential of adult human mesenchymal stem cells. *Science.* 1999;284(5411):143-7.
97. Filardo G, Perdisa F, Roffi A, Marcacci M, Kon E. Stem cells in articular cartilage regeneration. *J Orthop Surg Res.* 2016;11:42.
98. Pantalone A, Antonucci I, Guelfi M, Pantalone P, Usulli FG, Stuppia L, et al. Amniotic fluid stem cells: an ideal resource for therapeutic application in bone tissue engineering. *Eur Rev Med Pharmacol Sci.* 2016;20(13):2884-90.
99. Drengk A, Zapf A, Stürmer EK, Stürmer KM, Frosch KH. Influence of platelet-rich plasma on chondrogenic differentiation and proliferation of chondrocytes and mesenchymal stem cells. *Cells Tissues Organs.* 2009;189(5):317-26.
100. Zaky SH, Ottonello A, Strada P, Cancedda R, Mastrogiacomo M. Platelet lysate favours *in vitro* expansion of human bone marrow stromal cells for bone and cartilage engineering. *J Tissue Eng Regen Med.* 2008;2(8):472-81.
101. Wang Z, Hu H, Li Z, Weng Y, Dai T, Zong C, et al. Sheet of osteoblastic cells combined with platelet-rich fibrin improves the formation of bone in critical-size calvarial defects in rabbits. *Br J Oral Maxillofac Surg.* 2016;54(3):316-21.
102. Qiu G, Shi Z, Xu HHK, Yang B, Weir MD, Li G, et al. Bone regeneration in minipigs via calcium phosphate cement scaffold delivering autologous BMSCs and platelet-rich plasma. *J Tissue Eng Regen Med.* 2017.
103. Wei B, Huang C, Zhao M, Li P, Gao X, Kong J, et al. Effect of Mesenchymal Stem Cells and Platelet-Rich Plasma on the Bone Healing of Ovariectomized Rats. *Stem Cells Int.* 2016;2016:9458396.

104. Li H, Sun S, Liu H, Chen HUA, Rong XIN, Lou J, et al. Use of a biological reactor and platelet-rich plasma for the construction of tissue-engineered bone to repair articular cartilage defects. *Exp Ther Med.* 2016;12(2):711-19.
105. Tobita M, Tajima S, Mizuno H. Adipose tissue-derived mesenchymal stem cells and platelet-rich plasma: stem cell transplantation methods that enhance stemness. *Stem Cells Res Ther.* 2015;6:215.
106. Zuk PA, Zhu M, Mizuno H, Huang J, Futrell JW, Katz AJ, et al. Multilineage cells from human adipose tissue: implications for cell-based therapies. *Tissue Eng.* 2001;7(2):211-28.
107. Mizuno H, Zuk PA, Zhu M, Lorenz HP, Benhaim P, Hedrick MH. Myogenic differentiation by human processed lipoaspirate cells. *Plast Reconstr Surg.* 2002;109(1):199-209.
108. Puissant B, Barreau C, Bourin P, Clavel C, Corre J, Bousquet C, et al. Immunomodulatory effect of human adipose tissue-derived adult stem cells: Comparison with bone marrow mesenchymal stem cells. *Br J Haematol.* 2005;129(1):118-29.
109. Hicok KC, Du Laney TV, Zhou YS, Halvorsen Y-DC, Hitt DC, Cooper LF, et al. Human adipose-derived adult stem cells produce osteoid in vivo. *Tissue Eng.* 2004;10(3-4):371-80.
110. Miranville A, Heeschen C, Sengenès C, Curat CA, Busse R, Bouloumié A. Improvement of postnatal neovascularization by human adipose tissue derived stem cells. *Circulation.* 2004;110(3):349-55.
111. Mizuno H, Tobita M, Uysal AC. Adipose-derived stem cells as a novel tool for future regenerative medicine. *Stem Cells.* 2012;30(5):804-10.
112. Boquest AC, Shahdadfar A, Brinchmann JE, Collas P. Isolation of stromal stem cells from human adipose tissue. *Methods Mol Biol.* 2006;325:35-46.
113. Sze SK, de Kleijn DP, Lai RC, Khia Wey Tan E, Zhao H, Yeo KS, et al. Elucidating the secretion proteome of human embryonic stem cell-derived mesenchymal stem cells. *Mol Cell Proteomics.* 2007;6(10):1680-9.
114. Park BS, Jang KA, Sung JH, Park JS, Kwon YH, Kim KJ, et al. Adipose-derived stem cells and their secretory factors as a promising therapy for skin aging. *Dermatol Surg.* 2008;34(10):1323-6.
115. Caplan AI, Correa D. The MSC: an injury drugstore. *Cell Stem Cell.* 2011;9(1):11-5.
116. Shabbir A, Zisa D, Lin H, Mastri M, Roloff G, Suzuki G, et al. Activation of host tissue trophic factors through JAK-STAT3 signaling: a mechanism of mesenchymal stem cell-mediated cardiac repair. *Am J Physiol Heart Circ Physiol.* 2010;299(5):H1428-38.
117. Kocaoemer A, Kern S, Klüter H, Bieback K. Human AB serum and thrombin-activated platelet-rich plasma are suitable alternatives to fetal calf serum for the expansion of mesenchymal stem cells from adipose tissue. *Stem Cells.* 2007;25(5):1270-8.
118. Zhang Y, He J, Xiao G, Li Q. Effect of platelet-rich plasma on the proliferation and adipogenic differentiation of human adipose-derived stem cells in vitro. *Nan Fang Yi Ke Da Xue Xue Bao.* 2011;31(3):525-8.
119. Li H, Liu D, Yu Y, Wu T. Experimental research of the promotion effect of autogeneic PRP on osteogenic differentiation of human adipose-derived stem cells in vitro. *Zhongguo Xiu Fu Chong Jian Wai Ke Za Zhi.* 2009;23(6):732-6.
120. Russo A, Condello V, Madonna V, Guerriero M, Zorzi C. Autologous and micro-fragmented adipose tissue for the treatment of diffuse degenerative knee osteoarthritis. *J Exp Orthop.* 2017;4(1):33.
121. Feng Z, Liu J, Shen C, Lu N, Zhang Y, Yang Y, et al. Biotin-avidin mediates the binding of adipose-derived stem cells to a porous  $\beta$ -tricalcium phosphate scaffold: Mandibular regeneration. *Exp Ther Med.* 2016;11(3):737-46.
122. Yun S, Ku S, Kwon Y. Adipose-derived mesenchymal stem cells and platelet-rich plasma synergistically ameliorate the surgical-induced osteoarthritis in Beagle dogs. *J Orthop Surg Res.* 2016;11:9.
123. Cvetković VJ, Najdanović JG, Vukelić-Nikolić M, Stojanović S, Najman SJ. Osteogenic potential of in vitro osteo-induced adipose-derived mesenchymal stem cells combined with platelet-rich plasma in an ectopic model. *Int Orthop.* 2015;39(11):2173-80.
124. Tajima S, Tobita M, Orbay H, Hyakusoku H, Mizuno H. Direct and indirect effects of a combination of adipose-derived stem cells and platelet-rich plasma on bone regeneration. *Tissue Eng Part A.* 2015;21(5-6):895-905.
125. Van Pham P, Bui KH-T, Ngo DQ, Vu NB, Truong NH, Phan NL, et al. Activated platelet-rich plasma improves adipose-derived stem cell transplantation efficiency in injured articular cartilage. *Stem Cell Res Ther.* 2013;4(4):91.
126. De Bari C, Dell'Accio F, Tylzanowski P, Luyten FP. Multipotent mesenchymal stem cells from adult human synovial membrane. *Arthritis Rheum.* 2001;44(8):1928-42.
127. Tang H, Chen W, Chiang C, Chen L, Chang Y. Differentiation Effects of Platelet-Rich Plasma Concentrations on Synovial Fluid Mesenchymal Stem Cells from Pigs Cultivated in Alginate Complex Hydrogel. *Int J Mol Sci.* 2015;16(8):18507-21.
128. Dominici M, Le Blanc K, Mueller I, Slaper-Cortenbach I, Marini F, Krause D, et al. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy.* 2006;8(4):315-7.
129. Sakaguchi Y, Sekiya I, Yagishita K, Muneta T. Comparison of human stem cells derived from various mesenchymal tissues: Superiority of synovium as a cell source. *Arthritis Rheum.* 2005;52(8):2521-9.
130. Jones EA, Crawford A, English A, Henshaw K, Mundy J, Corscadden D, et al. Synovial fluid mesenchymal stem cells in health and early osteoarthritis: Detection and functional evaluation at the single-cell level. *Arthritis Rheum.* 2008;58(6):1731-40.
131. Sekiya I, Ojima M, Suzuki S, Yamaga M, Horie M, Koga H, et al. Human mesenchymal stem cells in synovial fluid increase in the knee with degenerated cartilage and osteoarthritis. *J Orthop Res.* 2012;30(6):943-9.
132. Morito T, Muneta T, Hara K, Ju YJ, Mochizuki T, Makino H, et al. Synovial fluid-derived mesenchymal stem cells increase after intra-articular ligament injury in humans. *Rheumatology (Oxford).* 2008;47(8):1137-43.
133. Matsukura Y, Muneta T, Tsuji K, Koga H, Sekiya I. Mesenchymal stem cells in synovial fluid increase after meniscus injury. *Clin Orthop Relat Res.* 2014;472:1357-64.
134. Lee J, Min HJ, Park HJ, Lee S, Seong SC, Lee MC. Synovial membrane-derived stem cells supported by Platelet-Rich Plasma can repair osteochondral defects in a rabbit model. *Arthroscopy.* 2013;29(6):1034-46.
135. Huang S, Wang Z. Influence of platelet-rich plasma on proliferation and osteogenic differentiation of skeletal muscle satellite cells: An in vitro study. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2010;110(4):453-62.
136. Mauro A. Satellite cell of skeletal muscle fibers. *J Biophys Biochem Cytol.* 1961;9:493-5.
137. Li H, Usas A, Poddar M, Chen CW, Thompson S, Ahani B, et al. Platelet-Rich Plasma Promotes the Proliferation of Human Muscle Derived Progenitor Cells and Maintains Their Stemness. *PLoS One.* 2013;8(6):e64923.
138. Cao B, Huard J. Muscle-derived stem cells. *Cell Cycle.* 2004;3(2):104-7.
139. Zheng B, Cao B, Crisan M, Sun B, Li G, Logar A, et al. Prospective identification of myogenic endothelial cells in human skeletal muscle. *Nat*

- Biotechnol. 2007;25(9):1025-34.
140. Crisan M, Yap S, Casteilla L, Chen CW, Corselli M, Park TS, et al. A perivascular origin for mesenchymal stem cells in multiple human organs. *Cell Stem Cell*. 2008;3(3):301-13.
141. Huang S, Jia S, Liu G, Fang D, Zhang D. Osteogenic differentiation of muscle satellite cells induced by platelet-rich plasma encapsulated in three-dimensional alginate scaffold. *Oral Surg Oral Med Oral Pathol Oral Radiol*. 2012;114(5):S32-S40.
142. Souza TF, Sakamoto SS, Ferreira GT, Gameiro R, Marinho M, de Andrade AL, et al. Osteogenic potential of mesenchymal cells derived from canine umbilical cord matrix co-cultured with platelet-rich plasma and demineralized bone matrix. *J Vet Sci*. 2015;16(3):381-4.
143. Cardoso TC, Ferrari HF, Garcia AF, Novais JB, Silva-Frade C, Ferrarezi MC, et al. Isolation and characterization of Wharton's jelly-derived multipotent mesenchymal stromal cells obtained from bovine umbilical cord and maintained in a defined serum-free three-dimensional system. *BMC Biotechnology*. 2012;12:18.
144. Filioli Uranio M, Valentini L, Lange-Consiglio A, Caira M, Guaricci AC, L'Abbate A, et al. Isolation, proliferation, cytogenetic, and molecular characterization and in vitro differentiation potency of canine stem cells from foetal adnexa: a comparative study by amniotic fluid, amnion, and umbilical cord matrix. *Mol Reprod Dev*. 2011;78(5):361-73.
145. Handschel J, Naujoks C, Langenbach F, Berr K, Depprich RA, Ommerborn MA, et al. Comparison of ectopic bone formation of embryonic stem cells and cord blood stem cells in vivo. *Tissue Eng Part A*. 2010;16(8):2475-83.
146. Park SB, Seo MS, Kim HS, Kang KS. Isolation and characterization of canine amniotic membrane-derived multipotent stem cells. *PLoS One*. 2012;7(9):e44693.
147. Seo MS, Jeong YH, Park JR, Park SB, Rho KH, Kim HS, et al. Isolation and characterization of canine umbilical cord blood-derived mesenchymal stem cells. *J Vet Sci*. 2009;10(3):181-7.
148. Lu L, Shen RN, Broxmeyer HE. Stem cells from bone marrow, umbilical cord blood and peripheral blood for clinical application: current status and future application. *Crit Rev Oncol Hematol*. 1996;22(2):61-78.
149. Wen Y, Gu W, Cui J, Yu M, Zhang Y, Tang C, et al. Platelet-rich plasma enhanced umbilical cord mesenchymal stem cells-based bone tissue regeneration. *Arch Oral Biol*. 2014;59(11):1146-54.
150. Prusa AR, Hengstschläger M. Amniotic fluid cells and human stem cell research: a new connection. *Med Sci Monit*. 2002;8(11):253-257.
151. Prusa AR, Marton E, Rosner M, Bernaschek G, Hengstschläger M. Oct-4-expressing cells in human amniotic fluid: a new source for stem cell research? *Hum Reprod*. 2003;18(7):1489-93.
152. Fauza D. Amniotic fluid and placental stem cells. *Best Pract Res Clin Obstet Gynaecol*. 2004;18(6):877-91.
153. De Coppi P, Bartsch G, Siddiqui MM, Xu T, Santos CC, Perin L, et al. Isolation of amniotic stem cell lines with potential for therapy. *Nat Biotechnol*. 2007;25(1):100-6.
154. In't Anker PS, Scherjon SA, Kleijburg-van der Keur C, Noort WA, Claas FH, Willemze R, et al. Amniotic fluid as a novel source of mesenchymal stem cells for therapeutic transplantation. *Blood*. 2003;102(4):1548-9.
155. Holden C. Stem cells. Versatile stem cells without the ethical baggage? *Science*. 2007;315(5809):170.
156. Da Sacco S, De Filippo RE, Perin L. Amniotic fluid as a source of pluripotent and multipotent stem cells for organ regeneration. *Curr Opin Organ Transplant*. 2011;16(1):101-5.
157. Zhang S, Geng H, Xie H, Wu Q, Ma X, Zhou J, et al. The heterogeneity of cell subtypes from a primary culture of human amniotic fluid. *Cell Mol Biol Lett*. 2010;15(3):424-39.
158. Antonucci I, Pietro RD, Alfonsi M, Centurione MA, Centurione L, Sancilio S, et al. Human second-trimester amniotic fluid cells are able to create embryoid body-like structures in vitro and to show typical expression profiles of embryonic and primordial germ cells. *Cell Transplant*. 2014;23(12):1501-15.
159. Savickiene J, Treigyte G, Baronaite S, Valiuliene G, Kaupinis A, Valius M, et al. Human amniotic fluid mesenchymal stem cells from second- and third-trimester amniocentesis: differentiation potential, molecular signature, and proteome analysis. *Stem Cells Int*. 2015;2015:319238.
160. Wang KH, Kao AP, Chang CC, Lin TC, Kuo TC. Upregulation of Nanog and Sox-2 genes following ectopic expression of Oct-4 in amniotic fluid mesenchymal stem cells. *Biotechnol Appl Biochem*. 2015;62(5):591-7.
161. Kim J, Lee Y, Kim H, Hwang KJ, Kwon HC, Kim SK, et al. Human amniotic fluid-derived stem cells have characteristics of multipotent stem cells. *Cell Prolif*. 2007;40(1):75-90.
162. Pappa KI, Anagnou NP. Novel sources of fetal stem cells: where do they fit on the developmental continuum? *Regen Med*. 2009;4(3):423-33.
163. Tsai MS, Lee JL, Chang YJ, Hwang SM. Isolation of human multipotent mesenchymal stem cells from second-trimester amniotic fluid using a novel two-stage culture protocol. *Hum Reprod*. 2004;19(6):1450-6.
164. Antonucci I, Iezzi I, Morizio E, Mastrangelo F, Pantalone A, Mattioli-Belmonte M, et al. Isolation of osteogenic progenitors from human amniotic fluid using a single step culture protocol. *BMC Biotechnol*. 2009;9:9.
165. Antonucci I, Pantalone A, De Amicis D, D'Onofrio S, Stuppia L, Palka G, et al. Human amniotic fluid stem cells culture onto titanium screws: a new perspective for bone engineering. *J Biol Regul Homeost Agents*. 2009;23(4):277-9.
166. Chen Q, Xiao P, Chen JN, Cai JY, Cai XF, Ding H, et al. AFM studies of cellular mechanics during osteogenic differentiation of human amniotic fluid-derived stem cells. *Anal Sci*. 2010; 26(10):1033-7.
167. Hipp JA, Hipp JD, Atala A, Soker S. Ethanol alters the osteogenic differentiation of amniotic fluid-derived stem cells. *Alcohol Clin Exp Res*. 2010;34(10):1714-22.
168. Wang M, Li H, Si J, Dai J, Shi J, Wang X, et al. Amniotic fluid-derived stem cells mixed with platelet rich plasma for restoration of rat alveolar bone defect. *Acta Biochim Biophys Sin (Shanghai)*. 2017;49(3):197-207.