



Aflatoxin B₁ Inhibits the Replication of Chandipura Virus in Human Hepatoma Cells

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Abstract

We previously showed that aflatoxin B₁ inhibited the antiviral type I interferon signalling response pathway as a mechanism of hepatocellular carcinoma. The type I interferon signalling is part of the innate immune system that defends the body against invading microorganisms. The consequence of compromising or suppressing the immune system include increased susceptibility to infectious diseases, reactivation of chronic diseases as well as increased susceptibility to cancers. Therefore, the chandipura virus was selected as a model virus to test the hypothesis that aflatoxin B₁ would enhance viral replication in patients presenting with viral infections and long exposure to dietary aflatoxin. The chandipura viral titre was quantified in the presence or absence of aflatoxin B₁ by plaque assay using L929 cells and stained with crystal violet. The data presented in this research notes is a side product of another research of which the results have been published. The current study showed aflatoxin B₁ (10 µM) reduced the replication of chandipura virus and the rate of reduction was exacerbated in the presence of interferon. The results open avenues for further studies to understand the interplay between viral infections and aflatoxins in accelerating cancers such as the hepatocellular carcinoma.

Keywords: Aflatoxin B₁; Chandipura virus; HepG2; Type I interferon pathway

Introduction

Aflatoxin B₁ (AFB₁) a lethal mycotoxin produced by *Aspergillus flavus* and *Aspergillus parasiticus* has been reported to inhibit the type I IFN response signalling pathway [1]. The type I IFN signalling response pathway is part of the innate immune system that is responsible for combating disease causing microorganisms and also initiate the process for the development of the adaptive immune response. The consequence of compromising the immune system include increased susceptibility to infectious diseases, reactivation of chronic diseases as well as increased susceptibility to cancers. What this suggests is that any agent that is capable of inhibiting the type I IFN response signalling pathway could significantly suppress the immune system and make the individual more prone to infections caused by microorganisms.

The current study aims to test the hypothesis that AFB₁ would enhance virus growth to higher titres in individuals with long term dietary exposure to aflatoxins. The data presented in this research notes is a side product of another research project of which the results have been published [1].

Materials and Methods

The human hepatoma cell line HepG2 (ECACC 85011430) and mouse fibroblast cells (L929) L929 (NCTC) (ECACC 85103115) were generously donated by Professor David J. Blackburn of the University of Surrey, UK. The cells were cultured in growth medium containing DMEM high glucose, L-glutamine, sodium pyruvate and HEPES (Sigma Aldrich, Germany) supplemented with 10% v/v heat inactivated Foetal Bovine Serum (FBS) (Sigma Aldrich, Germany), 1% v/v MEM non-essential amino acids (Sigma Aldrich, Germany), 100 IU/mL of penicillin and 100 µg/mL of streptomycin (Gibco life technologies, UK). The cultures were maintained at 37°C in 5% carbon dioxide (CO₂). In order to prevent mycoplasma infection and also to ensure that the cells were free of mycoplasma, occasionally actively dividing culture of cells were treated with 25 µg/mL of Plasmocin (cat no ant-mpt, invivogen, USA) an anti-mycoplasma agent. The lyophilized AFB₁ used in the study was commercially purchased from Sigma-Aldrich, USA (cat no A6636) and reconstituted as previously described [1]. The human interferon-alpha (IFN-α) used in the study was purchased from PBL interferon source (PBL, USA) and prepared as previously described [1]. The chandipura virus used in the study was generously donated by Professor David J. Blackburn.

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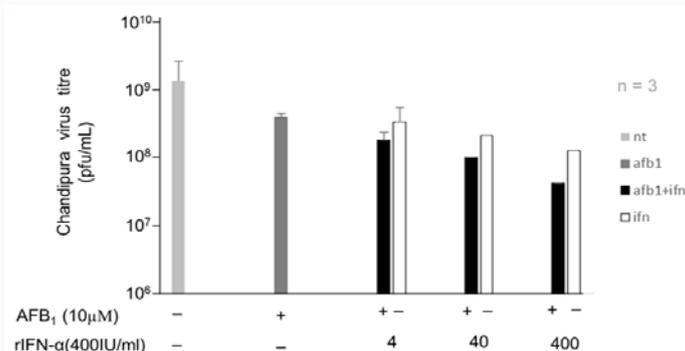


Figure 1: AFB₁ alone is lethal to chandipura virus and the lethality is exacerbated by IFN-α. Monolayers of HepG2 cells were pre-treated with AFB₁ (10µM) and simultaneously treated with increasing concentrations of rIFN-α (4, 40 & 400 IU/ml) for 24 hours. The cells were infected with chandipura virus at MOI of 0.1. The infected cells were incubated for one hour to allow the viruses to adsorb onto the cells. The virus containing media were removed and the cells treated with fresh media (without AFB₁ and or rIFN-α) for 24 hours. The tissue culture supernatants containing the virus were collected and the virus titre was quantified by plaque assay using L929 cells. After incubation for 3 days, plaques were identified by crystal violet staining and counted. The results are presented as mean and standard deviation of three independent experiments conducted in duplicate independent wells. Statistical analysis was performed using Microsoft excel (2013 version). NT: cells which were neither stimulated with interferon nor treated with aflatoxin B₁; AFB₁: cells which were treated with only aflatoxin B₁; AFB₁+IFN: cells which were stimulated with interferon and simultaneously treated with aflatoxin B₁; IFN: cells which were stimulated with only interferon.

To examine the effects of AFB₁ on replication of virus, chandipura virus was cultured in the presence or absence of AFB₁ (10 µM) and IFN-α (4, 40 and 400 IU/ml). The concentrations of AFB₁ and IFN-α used in this experiment were those reported previously [1]. Briefly, HepG2 cells were seeded at a density of 5×10^5 cells/well in 6-well plates. The cells were then stimulated with increasing concentration of IFN-α (4, 40 & 400 IU/ml) and simultaneously treated with or without AFB₁ (10 µM). Twenty-four hours later the cells were infected with chandipura virus at a Multiplicity of Infection (MOI) of 0.1. After 1 hour of incubation to allow the viruses to adsorb onto the surfaces of the cells, the supernatant was removed and 2 mL growth medium containing neither AFB₁ nor IFN-α were added. Cells were incubated for 24 hours and the supernatant collected and the virus titer determined by plaque assay using L929 cells following standard protocol. After incubation for three (3) days, plaques were stained with 0.05% crystal violet solution and counted. Titer concentrations (in plaque forming unit (PFU) per mL) were calculated by using the equation:

$$\text{PFU/mL} = \frac{\text{Average number of plaques}}{d \times v}$$

Where, d = dilution

v = volume of diluted virus added to the plate

Data were entered into excel spread sheet and Microsoft excel (2013 version) was used to calculate the averages and the standard deviations of the three independent experiments.

Results and Discussion

To be able to investigate the biological significance of the inhibition process on the anti-viral property of the type I IFN by AFB₁ in HepG2 cells, the survival of the HepG2 cells in the presence of AFB₁ was very critical. The concentration of AFB₁ employed in this study was 10 µM as previously reported [1]. This concentration was found to be non-toxic to the HepG2 cells but was high enough to significantly inhibit the type I IFN response signalling pathway. In addition, the concentration of IFN-α which induced maximally the type I IFN signalling was established as 400 IU/mL as previously reported [1]. Figure 1 shows AFB₁ reduced the replication of chandipura virus and the rate of reduction was exacerbated by activated anti-viral type I IFN response signalling pathway.

The type I IFN signalling pathway is well known for its antiviral property. Therefore, after demonstrating in a previous study that AFB₁ suppressed the type I IFN signalling pathway, the expectation was that AFB₁ would ultimately suppress that antiviral pathway and enhance viral replication to significantly higher titres.

A number of studies have utilized the replication of viruses to study the effects of different inhibitors or chemical groups on the antiviral effects of the type I IFN response signalling pathway. For example [2], utilized the replication of IFN sensitive Encephalomyocarditis Virus (EMCV) in vIRF-2 clone 3-9 and EV clone 5 cells to study the biological significance of vIRF-2 inhibition of the type I IFN signalling pathway and reported that vIRF-2 rescued the EMCV from the antiviral effects of type I IFN (2). In another study, [3] also employed the replication of Enterovirus in human cervical carcinoma (HeLa S3), Vero and human hepatoma (Huh 7) cell lines to determine whether proteinase 2 A^{Pro} was essential for Enterovirus replication in type I IFN treated cells and reported that proteinase 2 A^{Pro} rescued the Enterovirus from the antiviral effects of the type I IFN. However, proteinase 2 A^{Pro} could not rescue the replication of vesicular stomatitis virus and EMCV in type I IFN pre-treated cells [3].

Taken together the above information, the replication of chandipura virus was utilized in this study to investigate the biological significance of the AFB₁ inhibition of type I IFN signalling pathway in HepG2 cells. Contrary to the initial expectation that AFB₁ would rescue the IFN sensitive virus from the antiviral effects of the type I IFN response signalling pathway, our result pointed to the fact that AFB₁ rather significantly reduced the IFN-sensitive chandipura virus replication and that the rate of reduction was exacerbated by activated type I IFN response signalling pathway. The result obtained from this study was similar to that of [4] who reported that AFB₁ decreased the replication of hepatitis B virus (HBV) in HepaRG cells after exposure of HBV to AFB₁ at a concentration of up to 5 µM [4].

However, a study conducted by [5] to investigate the effects of AFB₁ exposure on the replication of duck hepatitis B virus in Pekin duck model during the initial stage of virus-AFB₁ interaction showed a significant increase in the titre of duck hepatitis B virus in the serum and liver of AFB₁ treated-ducks compared with Dimethyl Sulfoxide (DMSO) treated controls [5]. The increase in viral replication

observed in the Pekin duck model was confirmed *in vitro* in primary duck hepatocytes [5]. In addition, [6] investigated the effects of AFB₁ exposure on the replication of influenza virus in LLC-MK₂ cells and reported that the levels of virus concentration attained in AFB₁ treated cells were two to four-fold higher compared with the untreated control. The enhanced influenza viral growth rate observed in AFB₁ treated cells in the study of [6] was attributed to the ability of AFB₁ to inhibit IFN production [6].

There seem to be conflicting information in literature as to the effects of AFB₁ on the replication of viruses. While some studies have reported that AFB₁ enhances the replication of some viruses in the presence of the activated type I IFN response signalling pathway, others have reported that AFB₁ inhibits the replication of some viruses. Findings from this study indicates that AFB₁ reduced the replication of chandipura virus. The mechanism by which AFB₁ kills some viruses and rescues others in the presence of the activated type I IFN signalling pathway is a subject of further research. It is possible that the inability of AFB₁ to rescue some viruses but others could be due to the fact that the replication of some viruses may differ from others in their sensitivity to AFB₁ and the diverse activity of AFB₁ under experimental conditions. It is also possible that AFB₁ may bind and block certain enzymes and substrates that are required for the replication of some viruses hence its ability to kill them.

Conclusion

In conclusion the current study has shown that AFB₁ reduced the replication of chandipura virus and the rate of reduction was exacerbated in the presence of IFN-. Considering finding(s) from this and other studies, further research may be required to unravel the exact mechanism by which AFB₁ kills some viruses and rescues others.

Limitation(s)

The chandipura virus used in this study has tropism for cells of the nervous system and that its use on human liver derived cell line could

have affected the result. In place of chandipura virus, hepatitis B virus which has tropism for liver cells could have been used. However, the difficulty in procuring the virus made it impossible for it to be used in the current study.

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References

1. Narkwa PW, Blackbourn DJ, Mutocheluh M. Aflatoxin B1 inhibits the type I interferon response pathway via STAT1 suggesting another mechanism of hepatocellular carcinoma. *Infect Agent Cancers*. 2017;12:17.
2. Mutocheluh M, Hindle L, Aresté C, Chanas SA, Butler LM, Lowry K, et al. Kaposi's sarcoma-associated herpesvirus viral interferon regulatory factor-2 inhibits type 1 interferon signalling by targeting interferon-stimulated gene factor-3. *J Gen Virol*. 2011;92(Pt 10):2394-8.
3. Morrison JM, Racaniello VR. Proteinase 2Apro Is Essential for Enterovirus Replication in Type I Interferon-Treated Cells. *J Virol*. 2009;83(9):4412-22.
4. Lereau M, Gouas D, Villar S, Besaratinia A, Hautefeuille A, Berthillon P, et al. Interactions between hepatitis B virus and aflatoxin B1: effects on p53 induction in HepaRG cells. *J Gen Virol*. 2012;93:640-50.
5. Barraud L, Guerret S, Chevallier M, Borel C, Jamard C, Trepo C, et al. Enhanced Duck Hepatitis B Virus Gene Expression Following Aflatoxin B1 Exposure. *Hepatology*. 1999;29(4):1317-23.
6. Hahon N, Booth JA, Stewart JD. Aflatoxin inhibition of viral interferon induction. *Antimicrob Agents Chemother*. 1979;16(3):277-82.