



Immunological Response of Recombinant Vaccinia Virus Including the HIV-1 Gene

Takuma Hayashi* and Hiroshi Shibuta

Department of Infectious Diseases, University of Tokyo, Japan

Abstract

We examined mouse immune response to 4 kinds of recombinant vaccinia viruses carrying the human immunodeficiency virus type 1 (HIV-1) Group-specific Antigen(gag) gene, including vac-gag/pol, which produces HIV-1-like particles with processed gag proteins; vac-gag, which also produces HIV-1-like particles but with unprocessed gag protein; and vac-gag-pol-fuse and vac-es-gag/pol, neither of which produces such virus particles but releases Reverse Transcriptase (RT) and gag protein, respectively, from infected cells. Although infection of mice with recombinant vaccinia viruses induced production of the anti-HIV-1 gag protein 24 (p24) antibodies in all mice, vac-gag/pol and vac-es-pol induced higher production than the other two recombinants. Increase in thymidine [3H] uptake by splenic lymphocytes following HIV-1p24 antigen stimulation was most evident in mice infected with vac-gag/pol. Thus, the highest immune response, both humoral and cellular, was elicited by vac-gag/pol, indicating that among those tested; this recombinant vaccinia virus is the best candidate for a vaccine that induces anti-HIV-1 gag immunity.

Introduction

The Human Immunodeficiency Virus type 1 (HIV-1) envelope antigen (env) has been a primary target for the development of vaccines against HIV-1, the causative agent of Acquired Immune Deficiency Syndrome (AIDS). Previous infectious studies showed that recombinant vaccinia viruses constructed as vaccine candidates evoked humoral and cell-mediated immune reactions against various pathogens [1-4]. There are reports that recombinant vaccinia viruses carrying the env region of HIV-1 are capable of inducing antibodies against the env protein in mice and chimpanzees [5,6]. Furthermore, Zarling, et al confirmed the presence of T-cell responses to the env antigen in macaques immunized with a recombinant virus carrying the env gene [7]. However, it is known that the mutation rate of the env gene is high, and it has also been reported that immunity against HIV-1 glycoprotein 120 (gp120) of the env protein elicited by HIV-1 infection exerted deleterious effects on the immune system [8-10]. These findings suggest problems to be overcome before using the env protein as the target for vaccines.

Recently, attention has been paid to the HIV-1 Group-specific Antigen (gag) including viral core antigen because of the low rate of mutation in the gag region and specific decreases in anti-gag antibodies before the development of AIDS in HIV-1-infected patients [11,12]. Moreover, cellular immunity against viral core antigens is known to play a definitive role in recovery from viral infections [13], and several reports have indicated that cellular immunity against gag proteins was detectable in HIV-1 infection [14-16]. The primary product of the gag gene is the p55 gag precursor, which is processed into p17, p24 and p15 mature gag proteins by the protease encoded by the polymerase (pol) gene [14]. Several workers have already succeeded in expressing HIV-1 gag proteins in mammalian cells using recombinant vaccinia vectors [17-20] and recent reports showed that recombinant vaccinia viruses carrying the gag gene can produce HIV-1 like particles in infected mammalian cells [17-20]. However, only a little is known about the immunological potency of these recombinant vaccinia viruses. In the present study we addressed this matter using mice of 3 in bred strains as the host animal and 4 different kinds of recombinant vaccinia viruses carrying the HIV-1 gag gene as the immunogen. The recombinants included 3 previously described and one newly constructed [11].

Although several studies on the expression of the HIV-1 gag gene in recombinant vaccinia viruses have been reported, only a report described the immunological potency of the recombinants [17-21]. They constructed 5 recombinants, in which v-gag 1 and v-gag 4 were similar to our vac-gag/pol and vac-gag, respectively. C57/B6 mice immunized with these recombinants at a single

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*Correspondence:

Takuma Hayashi, Department of Infectious Diseases, Institution for Medical Science, University of Tokyo, Shiroganedai, Minato-ku, Tokyo, Japan, E-mail: yoyoyo224@hotmail.com

Received Date: 01 Apr 2018

Accepted Date: 26 Apr 2018

Published Date: 01 May 2018

Citation:

Hayashi T, Shibuta H. Immunological Response of Recombinant Vaccinia Virus Including the HIV-1 Gene. *Am J Clin Microbiol Antimicrob.* 2018; 1(4): 1018.

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dose of 10^7 Plaque Formation Unit (PFU) per mouse by footpad inoculation mainly generated antibodies directed to p24 but occasionally those against p17 as well. Cellular immunity against the gag proteins was not examined in the mice. However, these authors found lymphoproliferative responses to psoralen-inactivated HIV-1 in chimpanzees immunized twice with a recombinant carrying the 3'-truncated gag gene (v-gag 5), each at a viral dose of 2×10^8 PFU, by skin scarification. In the present study we examined the immunogenic properties of our 4 recombinants carrying the HIV-1 gag gene. All the recombinants could effectively elicit anti-p24 antibody in mice of 3 inbred strains. Vac-gag/pol and vac-es-gag/pol seemed to be more effective than vac-gag and vac-gag-pol-fuse. Interestingly, there was no obvious correlation between the neutralizing antibody titer against vaccinia virus and that of anti-p24 antibody. In contrast to the production of anti-p24 antibody, splenic lymphocyte response to the p24 antigen, which was estimated as an indicator of cellular immunity, was most evident in the mouse group immunized with vac-gag/pol, irrespective of mouse strain, although the response index greatly varied from mouse to mouse, and a high viral dose of 10^8 PFU per mouse was necessary to induce the response. Vac-es-gag/pol also seemed to be effective in evoking the response, but statistically there was no significant difference between vac-es-gag/pol and vac-gag in all mouse strains. All the findings in the present study strongly suggest that vac-gag/pol is superior to the other recombinants carrying the gag gene in inducing immune responses, both humoral and cellular, against the gag protein. The most characteristic feature of vac-gag/pol is the capability to produce HIV-1-like particles composed of the processed gag proteins, which may well be correlated to the high immunological potency [19,20,22]. Vac-gag is also able to produce HIV-1-like particles, but its gag protein is the unprocessed precursor [19,20,23]. The newly constructed recombinant, vac-es/gag/pol, induced a fairly high production of anti-p24 antibody as well as considerable splenic lymphocyte response, which might correspond well to the efficient release of a modified gag protein from infected cells. However, this recombinant could not exceed vac-gag/pol in its immunological potency. We had expected that the sera from mice infected with these recombinants possessed neutralizing activity against HIV-1, since two mouse monoclonal antibodies against the p17 gag protein were reported to have the ability to neutralize HIV-1 [24,25]. However, our mouse sera failed to show this ability, and it remains to be examined whether these sera have antibody against the p17 gag protein. Although we measured here the splenic lymphocyte response to the p24 antigen as an indicator for evaluating cellular immunity, it is important to further clarify whether our recombinants, especially vac-gag/pol, are able to induce cytotoxic T cells against the cells expressing the gag antigen. If they have such ability, we could expect that they interfere with the onset of AIDS from asymptomatic HIV-1 carrier [26].

References

- Bennick JR, Yewdel LJW, Smith GL, Moller C, Moss B. Recombinant vaccinia virus primes and stimulates influenza haemagglutinin-specific cytotoxic T cells. *Nature*. 1984;311(5986):578-9.
- Panicali D, Davis SW, Weinberg RL, Paoletti E. Construction of live vaccines by using genetically engineered poxviruses: biological activity of recombinant vaccinia virus expressing influenza virus hemagglutinin. *Proc Natl Acad Sci USA*. 1983;80:5364-8.
- Paoletti E, Lipinskas BR, Samsonoff C, Mercer S, Panicali D. Construction of live vaccines using genetically engineered poxviruses: biological activity of vaccinia virus recombinants expressing the hepatitis B virus surface antigen and the herpes simplex virus glycoprotein D. *Proc Natl Acad Sci USA*. 1984;81(1):193-7.
- Smith GL, Mackett M, Moss B. Infectious vaccinia virus recombinants that express hepatitis B virus surface antigen. *Nature*. 1983;302(5908):490-5.
- Flu SL, Kosowski SG, Dalrymple JM. Expression of AIDS virus envelope gene in recombinant vaccinia viruses. *Nature*. 1986;320(6062): 537-40.
- Flu SL, Fultz PN, McClure HM, Eichberg JW, Thomas EK, Zarlring MJ, et al. Effect of immunization with a vaccinia-HIV env recombinant on HIV infection of chimpanzees. *Nature*. 1987;328(6132):721-3.
- Zarlring JM, Morton W, Moran PA, McClure J, Kosowski SG, Hu S-L. T-cell responses to human AIDS virus in macaques immunized with recombinant vaccinia viruses. *Nature*. 1986; 323(6086): 344-8.
- Siliciano RF, Lawton T, Knall C, Karr RW, Berman P, Gregory T, et al. Analysis of host-virus interactions in AIDS with anti-gp120 T cell clones: effect of HIV sequence variation and a mechanism for CD4+ cell depletion. *Cell*. 1988;54(4): 561-75.
- Lanzavecchia A, Roosnek E, Regory T, Berman P, Abrignani S. T cells can present antigens such as HIV gp120 targeted to their own surface molecules. *Nature*. 1988;334(6182): 530-2.
- Weinhold KJ, Lyerly HK, Matthews TJ, Tyler DS, Ahearne PM, Stine KC, et al. Cellular anti-gp120 cytolytic reactivities in HIV-1 seropositive individuals. *Lancet*. 1988;1(8591): 902-5.
- Shiipbach J, Haller O, Vogt M, Liithy R, Joller H, Oelz O, et al. Antibodies to HTIX-III in Swiss patients with AIDS and pre AIDS and in groups at risk for AIDS. *N Eng J Med*. 1985; 312: 265-70.
- Weber JN, Clapham PR, Weiss RA, Parker D, Roberts C, Duncan J, et al. Human immunodeficiency virus infection in two cohorts of homosexual men: neutralizing sera and association of anti-gag antibody with prognosis. *Lancet*. 1987;1(8525):19-22.
- Yewdel JW, Bennink JR, Smith GL, Moss B. Influenza A virus nucleoprotein is a major target antigen for cross reactive anti-influenza A virus cytotoxic T lymphocytes. *Proc Natl Acad Sci USA*. 1985;82(6):1785-9.
- Nixon DF, Townsend ARM, Elvin JG, Rizza CR, Gallwey J, McMichael AJ. HIV-1 gag-specific cytotoxic T lymphocytes defined with recombinant vaccinia virus and synthetic peptides. *Nature*. 1988;336(6198): 484-7.
- Plata F, Autran B, Martins LP, Wain-Hobson S, Raphael M, Mayaud C, et al. AIDS virus-specific cytotoxic T lymphocytes in lung disorders. *Nature*. 1987; 328(6128):348-51.
- Walker BD, Chakrabarti S, Moss B, Paradis TJ, Flynn T, Durno AG, et al. HIV-specific cytotoxic T lymphocytes in seropositive individuals. *Nature*. 1987;328(6128):345-8.
- Karacostas V, Nagashima K, Gonda MA, Moss B. Human immunodeficiency virus-like particles produced by a vaccinia virus expression vector. *Proc Natl Acad Sci USA*. 1989;86(22):8964-7.
- Shioda T, Shibuta H. Production of human immunodeficiency virus (HIV)-like particles from cells infected with recombinant vaccinia viruses carrying the gag gene of HIV. *Virology*. 1990;175(1):139-48.
- Hayashi T, Shioda T, Iwakura Y, Shibuta H. RNA packaging signal of human immunodeficiency virus type 1. *Virology*. 1992;188(2): 590-9.
- Hayashi T, Ueno Y, Okamoto T. Elucidation of a conserved RNA stem-loop structure in the packaging signal of human immunodeficiency virus type 1. *FEBS Lett*. 1993;327(2): 213-8.
- Mak TK, Hesselring AC, Hussey GD, Cotton MF. Making BCG vaccination programmes safer in the HIV era. *Lancet*. 2008;372(9641):786-7.
- Klein F, Mouquet H, Dosenovic P, Scheid JF, Scharf L, Nussenzweig MC. Antibodies in HIV-1 vaccine development and therapy. *Science*. 2013;341(6151):1199-204.

23. Hayden EC. Almighty antibodies? A new wave of antibody-based approaches aims to combat HIV. *Nat Med.* 2015;21(7):657-9.
24. Papsidero LD, Sheu M, Ruscetti FW. Human immunodeficiency virus type 1-neutralizing monoclonal antibodies which react with p17 core protein: characterization and epitope mapping. *J Virol.* 1989;63:267-72.
25. Thippeshappa R, Tian B, Cleveland B, Guo W, Polacino P, Hu SL. Oral Immunization with Recombinant Vaccinia Virus Prime and Intramuscular Protein Boost Provides Protection against Intra-Rectal SHIV Challenge in Macaques. *Clin Vaccine Immunol.* 2016;23(3):204-12.
26. Kannagi NI, Masuda T, Hattori T, Kanoh T, Nasu K, Yamamoto, et al. Interference with human immunodeficiency virus (HIV) replication by CD8+ T cells in peripheral blood leukocytes of asymptomatic HIV carriers *in vitro.* *J Virol.* 1990; 64(7):3399-406.