



# Citrullinated Proteins are Arthritogenic Autoantigens in Rheumatoid Arthritis

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## Abstract

Both genetic and environmental factors are implicated for the development of Rheumatoid Arthritis (RA). The association of RA with HLA-DRB1 shared epitope (SE) suggests that T cells recognize an antigen presented on HLA-DRB1SE allele. The discovery of Anti Citrullinated Peptide Antibodies (ACPAs) greatly advanced our understanding of RA pathogenesis. ACPAs, including ACPAs against peptides derived from Epstein-Barr Virus (EBV), appear many years before the onset of clinical RA and are associated with severe disease. Citrullination is the conversion of arginine residues to citrulline in proteins, mediated by Peptidyl Arginine Deiminases (PADs). Citrullination increases the peptide binding affinity to HLA-DRB1SE that can activate T cells and provide help to B cells for ACPA production and there is increased frequency of T cells recognizing citrullinated (cit) peptides in ACPA + RA. Periodontal Disease (PD), a risk factor for RA, can provide cit neoantigens. *Porphyromonas gingivalis*, a causative agent for PD, expresses PAD (PPAD), and arginine ginpains (Rgps) which preferentially cleave proteins at terminal arginine residues. Cross-reactivity was detected between ACPAs against *P. gingivalis* or EBV peptides and human peptides which can re-direct immune response to self-antigens. There is some evidence suggesting that ACPAs are arthritogenic. In animal models, cit peptides can cause or exacerbate arthritis in a HLA-DRB1SE-restricted manner. ACPAs can also activate macrophages and cause bone resorption.

**Keywords:** Anti-cyclic citrullinated antibodies; Autoantigen; Autoantibody; Rheumatoid arthritis

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## Introduction

The etiology of Rheumatoid Arthritis (RA) is incompletely understood. Twin studies have shown that environmental factors rather than genetic factors appear to play a major role in the development of the disease [1]. Periodontal Disease (PD) and cigarette smoking are two environmental risk factors for RA. In the last ten or so years progress has been made that advanced our understanding on RA induction and this will be reviewed in this article.

## Rheumatoid Arthritis is an Antigen-Driven Immune Disease

In the 1970s RA has been found to be associated with HLA-DR4, and this association has been expanded later to other HLA-DR alleles. All HLA-DR alleles associated with RA bear a common DR β chain sequence at positions 70-74, known as HLA-DRB1 shared epitope (HLA-DRB1\*SE) [2,3]. The function of HLA-DRB1 molecules is to present antigenic peptides to T cells via the T cell antigen receptor (TCR). Therefore, this HLA-DR association plus the heavy infiltration of arthritic synovial membrane with macrophages and T cells and the increased levels of inflammatory T cell-cytokines in arthritic joints suggested that RA is a T cell-mediated disease. However, the inciting antigen was not known. The involvement of B cells attracted much less attention. Yet, patients with RA have Rheumatoid Factor (RF), an autoantibody against the Fc region of IgG, but the pathogenetic role of RF remained elusive. In RA synovial membrane there are ectopic lymphoid structures with features of germinal center and ability to present antigen and differentiate B cells into antibody-producing plasma cells. In early 2000s RA patients were found to have anti-citrullinated peptide antibodies (ACPAs) and this gave a strong push to efforts for elucidating the etiopathogenesis of RA.

## B-Cells Recognize Citrullinated Autoantigens: The Anti-Citrullinated Peptide Antibodies

ACPAs recognize citrullinated epitopes of various proteins. Citrullination is a post-translational

conversion of arginine to citrulline mediated by the enzymes arginine deiminases (PADs). ACPAs appear up to 12 years before clinical manifestations of RA and their frequency increases with approaching the onset of clinical RA [4,5]. In fact, these ACPAs, pre-dated the onset of RA are directed against endogenous citrullinated antigens and viral citrullinated antigens derived from Epstein-Barr virus nuclear antigen (EBNA1 and EBNA2) [6]. ACPAs appear in low levels and against one or few autoantigens at first, and their levels and specificities increase at the preclinical inflammatory phase of RA [7,8].

Cigarette smoking is a risk factor for ACPA+RA [9-11] and for PD [12]. In patients with RA, ACPAs are associated with HLA-DRB1\*SE and smoking [13,14]. More interestingly, in undifferentiated arthritis, HLA-DRB1SE is a risk factor for ACPA and not an independent risk factor for the development of RA [15]. There appear to be a dosage effect among HLA-DRB1SE alleles for ACPA(+) RA. Patients carrying both HLA-DRB1\*SE alleles had higher Odds Ratio for ACPA(+) RA than patients carrying just one HLA-DRB1\*SE allele [16]. There is also a significant interaction between cigarette smoking and HLA-DRB1\*SE for ACPA(+) RA [17-20]. Cigarette smoking is also a risk factor for RA development in pre-RA individuals [21].

ACPs are produced in RA arthritis joints. A high percentage (~25%) of IgG-expressing B cells from ACPA(+) RA synovial fluid recognize citrullinated autoantigens [22]. Furthermore, ACPA production may be triggered by Epstein-Barr Virus (EBV) infection. For instance, ectopic lymphoid structures from ACPA(+) RA synovial membrane transplanted into severe combined immunodeficiency (SCID) mice are able to sustain ACPA production plus anti-EBV antibodies [23].

Periodontal disease (PD) is associated with RA and this is attributed to *Porphyromonas gingivalis*, a causative agent for PD. *P. gingivalis* expresses two unique enzymes, peptidylarginine deiminase (PPAD) that citrullinates bacterial and human proteins, and arginine gingipains which are extracellular proteases cleaving proteins at arginine residues [24,25]. PPAD has specificity for C-terminal arginine and along with arginine gingipains can citrullinate many peptides from key RA autoantigens [26]. PAD and PPAD activities in periodontium are elevated in RA and non-RA patients with PD and this site may be the initial locus of ACPA production [27]. *Aggregatibacter actinomycetemcomitans*, a causative agent for PD, can cause citrullination of proteins via neutrophil membranes pore-forming leukotoxin-A, and there is evidence of *A. actinomycetemcomitans* infection in RA patients [28].

## T-Cells Recognize Citrullinated Proteins

The production of ACPAs most likely require T cell help, since ACPAs undergo isotype switching (which requires T cell help), and are associated with HLA-DRB1\*SE. Indeed, it has been shown that T cells recognize citrullinated peptides presented by HLA-DRB1\*SE. The conversion of arginine to citrulline at a peptide side chain that interacts with the P4 pocket (aa 71 of DR $\beta$  chain) of HLA-DRB1\*SE significantly increases HLA affinity and leads to activation of CD4+ T cells in DR4-IE transgenic mice [29]. Citrulline, not arginine, is accommodated in the electropositive P4 pocket of the HLA-DRB1\*04:01/04 [30]. Using HLA-II tetramers, Scally et al. [30] found citrullinated vimentin- and citrullinated aggrecan-specific CD4+ T cells in peripheral blood of HLA-DRB1\*04:01(+) RA patients. Naturally processed peptides from citrullinated vimentin were also

recognized by T cells in ACPA+, HLA-DRB1\*04:01(+) RA patients [31]. In mice, autophagy is key event involved in the generation of citrullinated peptides by antigen presenting cells to be presented to T cells [32].

## Infectious Agents may Trigger ACPA Production

Bacterial infection by *P. gingivalis*, may initiate the breaking of tolerance and autoimmunity in RA. ACPAs against the immunodominant  $\alpha$ -enolase peptide 1 (CEP1) cross-reacted with recombinant *P. gingivalis* enolase [33]. Furthermore, 20% of plasmablasts-derived antibodies from ACPA(+) RA patients were ACPAs and 63% of these ACPAs cross-reacted with outer membrane antigens and/or citrullinated enolase from *P. gingivalis* [34]. We already mentioned that EBV may trigger ACPA production in RA synovial membrane.

## Citrullinated Autoantigens are Pathogenic

ACPs are associated with extra-articular features of RA and disease severity and predict radiographic joint erosions [35-38]. Extracellular PAD levels in synovial fluid also correlate with disease activity in RA patients [39].

ACPs promote pro-inflammatory immune responses. Immune complexes of ACPA with fibrinogen induce macrophage TNF $\alpha$  production [40-42] via Fc $\gamma$ R and TLR4 [40]. ACPAs also activate complement. In fact, low avidity ACPAs are more potent activators of complement and are associated with higher rate of joint destruction [43]. RF potentiates the pro-inflammatory effects of ACPAs on complement. IgM and IgA RF amplify complement-dependent pro-inflammatory functions of ACPAs [44-46] and this has clinical ramifications. RF and ACPAs have additive effect on erosions, number and size, but RF augments erosion burden only in ACPA(+) RA patients [47]. In addition, ACPAs can promote pro-inflammatory responses through alteration of microRNAs (miRNAs). For instance, ACPAs reduced expression of let-7a miRNA in monocytic cell line and monocytes from ACPA(+) RA patients [48]. It should be noted that let-7a transfection of monocytic cell line decreases IL-1 $\beta$  expression [48].

Citrullinated peptides are arthritogenic in the context of HLA-DRB1SE background. In HLA-DRB1\*04:01 patients with RA a large proportion of peripheral blood T cells recognizing citrullinated peptides exhibit Th1 memory phenotype producing the pro-inflammatory IFN $\gamma$  [49]. Citrullinated fibrinogen induced arthritis in DR4-IE transgenic mice but not in wild-type C57BL/6 mice [50]. Furthermore, unmodified fibrinogen could not induce arthritis in DR4-IE transgenic mice [50]. However, recombinant human  $\alpha$ -enolase and *P. gingivalis* enolase either citrullinated or uncitrullinated induced arthritis in DR4-IE transgenic mice [51]. *P. gingivalis* exacerbated murine collagen-induced arthritis that was dependent on *P. gingivalis* PPAD [52]. On the other hand, a PAD inhibitor reduced severity of collagen-induced arthritis in mice [53]. ACPAs against fibrinogen also enhanced arthritis induced by monoclonal antibodies against collagen type II [54].

ACPs can also affect bone loss. For instance, human osteoclasts express PAD which induces vimentin citrullination during osteoclastogenesis. Furthermore, ACPAs directed against mutated citrullinated vimentin causes osteoclastogenesis and bone resorption both *in vitro* and *in vivo* [55]. Osteoclastogenesis and bone loss by

ACPAs are mediated through PAD enzymes and IL-8 *in vitro* and *in vivo* [56].

## References

- Sakkas LI, Bogdanos DP, Katsiari C, Platsoucas CD. Anti-citrullinated peptides as autoantigens in rheumatoid arthritis-relevance to treatment. *Autoimmun Rev.* 2014;13(11):1114-20.
- Gregersen PK, Silver J, Winchester RJ. The shared epitope hypothesis. An approach to understanding the molecular genetics of susceptibility to rheumatoid arthritis. *Arthritis Rheum.* 1987;30(11):1205-13.
- Wordsworth BP, Lanchbury JS, Sakkas LI, Welsh KI, Panayi GS, Bell JI. HLA-DR4 subtype frequencies in rheumatoid arthritis indicate that DRB1 is the major susceptibility locus within the HLA class II region. *Proc Natl Acad Sci U S A.* 1989;86(24):10049-53.
- Nielen MM, van Schaardenburg D, Reesink HW, van de Stadt RJ, van der Horst-Bruinsma IE, de Koning MH, et al. Specific autoantibodies precede the symptoms of rheumatoid arthritis: a study of serial measurements in blood donors. *Arthritis Rheum.* 2004;50(2):380-6.
- Arkema EV, Goldstein BL, Robinson W, Sokolove J, Wagner CA, Malspeis S, et al. Anti-citrullinated peptide autoantibodies, human leukocyte antigen shared epitope and risk of future rheumatoid arthritis: a nested case-control study. *Arthritis Res Ther.* 2013;15(5):R159.
- Johansson L, Pratesi F, Brink M, Arlestig L, D'Amato C, Bartaloni D, et al. Antibodies directed against endogenous and exogenous citrullinated antigens pre-date the onset of rheumatoid arthritis. *Arthritis Res Ther.* 2016;18(1):127.
- Sokolove J, Bromberg R, Deane KD, Lahey LJ, Derber LA, Chandra PE, et al. Autoantibody epitope spreading in the pre-clinical phase predicts progression to rheumatoid arthritis. *PLoS One.* 2012;7(5):e35296.
- van de Stadt LA, de Koning MH, van de Stadt RJ, Wolbink G, Dijkmans BA, Hamann D, et al. Development of the anti-citrullinated protein antibody repertoire prior to the onset of rheumatoid arthritis. *Arthritis Rheum.* 2011;63(11):3226-33.
- Too CL, Murad S, Hansson M, Alm LM, Dhaliwal JS, Holmdahl R, et al. Spectrum of Anti Citrullinated Peptide Antibodies (ACPA) fine specificities is different between Malaysian and Swedish rheumatoid arthritis patients: Implications for pathogenesis of RA. *Arthritis Rheumatol.* 2017;69(1):58-69.
- Lee HS, Irigoyen P, Kern M, Lee A, Batliwalla F, Khalili H, et al. Interaction between smoking, the shared epitope, and anti-cyclic citrullinated peptide: a mixed picture in three large North American rheumatoid arthritis cohorts. *Arthritis Rheum.* 2007;56(6):1745-53.
- van der Helm-van Mil AH, Verpoort KN, le Cessie S, Huizinga TW, de Vries RR, Toes RE. The HLA-DRB1 shared epitope alleles differ in the interaction with smoking and predisposition to antibodies to cyclic citrullinated peptide. *Arthritis Rheum.* 2007;56(2):425-32.
- Tomar SL, Asma S. Smoking-attributable periodontitis in the United States: findings from NHANES III. National Health and Nutrition Examination Survey. *J Periodontol.* 2000;71(5):743-51.
- Klareskog L, Stolt P, Lundberg K, Kallberg H, Bengtsson C, Grunewald J, et al. A new model for an etiology of rheumatoid arthritis: smoking may trigger HLA-DR (shared epitope)-restricted immune reactions to autoantigens modified by citrullination. *Arthritis Rheum.* 2006;54(1):38-46.
- Wagner CA, Sokolove J, Lahey LJ, Bengtsson C, Saevarsdottir S, Alfredsson L, et al. Identification of anticitrullinated protein antibody reactivities in a subset of anti-CCP-negative rheumatoid arthritis: association with cigarette smoking and HLA-DRB1 'shared epitope' alleles. *Ann Rheum Dis.* 2015;74(3):579-86.
- van der Helm-van Mil AH, Verpoort KN, Breedveld FC, Huizinga TW, Toes RE, de Vries RR. The HLA-DRB1 shared epitope alleles are primarily a risk factor for anti-cyclic citrullinated peptide antibodies and are not an independent risk factor for development of rheumatoid arthritis. *Arthritis Rheum.* 2006;54(4):1117-21.
- Balandraud N, Picard C, Reviron D, Landais C, Toussiroot E, Lambert N, et al. HLA-DRB1 genotypes and the risk of developing anti citrullinated protein antibody (ACPA) positive rheumatoid arthritis. *PLoS One.* 2013;8(5):e64108.
- Linn-Rasker SP, van der Helm-van Mil AH, van Gaalen FA, Kloppenburg M, de Vries RR, le Cessie S, et al. Smoking is a risk factor for anti-CCP antibodies only in rheumatoid arthritis patients who carry HLA-DRB1 shared epitope alleles. *Ann Rheum Dis.* 2006;65(3):366-71.
- Kallberg H, Padyukov L, Plenge RM, Ronnelid J, Gregersen PK, van der Helm-van Mil AH, et al. Gene-gene and gene-environment interactions involving HLA-DRB1, PTPN22, and smoking in two subsets of rheumatoid arthritis. *Am J Hum Genet.* 2007;80(5):867-75.
- Karlson EW, Chang SC, Cui J, Chibnik LB, Fraser PA, De Vivo I, et al. Gene-environment interaction between HLA-DRB1 shared epitope and heavy cigarette smoking in predicting incident rheumatoid arthritis. *Ann Rheum Dis.* 2010;69(1):54-60.
- Willemze A, van der Woude D, Ghiddey W, Levarht EW, Stoeken-Rijsbergen G, Verduyn W, et al. The interaction between HLA shared epitope alleles and smoking and its contribution to autoimmunity against several citrullinated antigens. *Arthritis Rheum.* 2011;63(7):1823-32.
- Fisher BA, Cartwright AJ, Quirke AM, de Pablo P, Romaguera D, Panico S, et al. Smoking, *Porphyromonas gingivalis* and the immune response to citrullinated autoantigens before the clinical onset of rheumatoid arthritis in a Southern European nested case-control study. *BMC Musculoskeletal Disord.* 2015;16:331.
- Amara K, Steen J, Murray F, Morbach H, Fernandez-Rodriguez BM, Joshua V, et al. Monoclonal IgG antibodies generated from joint-derived B cells of RA patients have a strong bias toward citrullinated autoantigen recognition. *J Exp Med.* 2013;210(3):445-55.
- Croia C, Serafini B, Bombardieri M, Kelly S, Humby F, Severa M, et al. Epstein-Barr virus persistence and infection of autoreactive plasma cells in synovial lymphoid structures in rheumatoid arthritis. *Ann Rheum Dis.* 2013;72(9):1559-68.
- Wegner N, Wait R, Sroka A, Eick S, Nguyen KA, Lundberg K, et al. Peptidylarginine deiminase from *Porphyromonas gingivalis* citrullinates human fibrinogen and alpha-enolase: implications for autoimmunity in rheumatoid arthritis. *Arthritis Rheum.* 2010;62(9):2662-72.
- Quirke AM, Lugli EB, Wegner N, Hamilton BC, Charles P, Chowdhury M, et al. Heightened immune response to autocitrullinated *Porphyromonas gingivalis* peptidylarginine deiminase: a potential mechanism for breaching immunologic tolerance in rheumatoid arthritis. *Ann Rheum Dis.* 2014;73(1):263-9.
- Montgomery AB, Kopec J, Shrestha L, Thezenas ML, Burgess-Brown NA, Fischer R, et al. Crystal structure of *Porphyromonas gingivalis* peptidylarginine deiminase: implications for autoimmunity in rheumatoid arthritis. *Ann Rheum Dis.* 2016;75(6):1255-61.
- Laugisch O, Wong A, Sroka A, Kantyka T, Koziel J, Neuhaus K, et al. Citrullination in the periodontium-a possible link between periodontitis and rheumatoid arthritis. *Clin Oral Investig.* 2016;20(4):675-83.
- Konig MF, Abusleme L, Reinholdt J, Palmer RJ, Teles RP, Sampson K, et al. Aggregatibacter actinomycetemcomitans-induced hypercitrullination links periodontal infection to autoimmunity in rheumatoid arthritis. *Sci Transl Med.* 2016;8(369):369ra176.
- Hill JA, Southwood S, Sette A, Jevnikar AM, Bell DA, Cairns E. Cutting edge: the conversion of arginine to citrulline allows for a high-affinity peptide interaction with the rheumatoid arthritis-associated HLA-DRB1\*0401 MHC class II molecule. *J Immunol.* 2003;171(2):538-41.

30. Scally SW, Petersen J, Law SC, Dudek NL, Nel HJ, Loh KL, et al. A molecular basis for the association of the HLA-DRB1 locus, citrullination, and rheumatoid arthritis. *J Exp Med*. 2013;210(12):2569-82.
31. Feitsma AL, van der Voort EI, Franken KL, el Bannoudi H, Elferink BG, Drijfhout JW, et al. Identification of citrullinated vimentin peptides as T cell epitopes in HLA-DR4-positive patients with rheumatoid arthritis. *Arthritis Rheum*. 2010;62(1):117-25.
32. Ireland JM, Unanue ER. Autophagy in antigen-presenting cells results in presentation of citrullinated peptides to CD4 T cells. *J Exp Med*. 2011;208(13):2625-32.
33. Lundberg K, Kinloch A, Fisher BA, Wegner N, Wait R, Charles P, et al. Antibodies to citrullinated alpha-enolase peptide 1 are specific for rheumatoid arthritis and cross-react with bacterial enolase. *Arthritis Rheum*. 2008;58(10):3009-19.
34. Li S, Yu Y, Yue Y, Liao H, Xie W, Thai J, et al. Autoantibodies From Single Circulating Plasmablasts React With Citrullinated Antigens and *Porphyromonas gingivalis* in Rheumatoid Arthritis. *Arthritis Rheumatol*. 2016;68(3):614-26.
35. Alexiou I, Germenis A, Ziogas A, Theodoridou K, Sakkas LI. Diagnostic value of anti-cyclic citrullinated peptide antibodies in Greek patients with rheumatoid arthritis. *BMC Musculoskelet Disord*. 2007;8:37.
36. Turesson C, Eberhardt K, Jacobsson LT, Lindqvist E. Incidence and predictors of severe extra-articular disease manifestations in an early rheumatoid arthritis inception cohort. *Ann Rheum Dis*. 2007;66(11):1543-4.
37. Alexiou I, Germenis A, Koutroumpas A, Kontogianni A, Theodoridou K, Sakkas LI. Anti-cyclic citrullinated peptide-2 (CCP2) autoantibodies and extra-articular manifestations in Greek patients with rheumatoid arthritis. *Clin Rheumatol*. 2008;27(4):511-3.
38. Jilani AA, Mackworth-Young CG. The role of citrullinated protein antibodies in predicting erosive disease in rheumatoid arthritis: a systematic literature review and meta-analysis. *Int J Rheumatol*. 2015;2015:728610.
39. Damgaard D, Senolt L, Nielsen CH. Increased levels of peptidylarginine deiminase 2 in synovial fluid from anti-CCP-positive rheumatoid arthritis patients: Association with disease activity and inflammatory markers. *Rheumatology (Oxford)*. 2016;55(5):918-27.
40. Clavel C, Nogueira L, Laurent L, Iobagiu C, Vincent C, Sebbag M, et al. Induction of macrophage secretion of tumor necrosis factor alpha through Fc gamma receptor IIa engagement by rheumatoid arthritis-specific autoantibodies to citrullinated proteins complexed with fibrinogen. *Arthritis Rheum*. 2008;58(3):678-88.
41. Sokolove J, Zhao X, Chandra PE, Robinson WH. Immune complexes containing citrullinated fibrinogen costimulate macrophages via Toll-like receptor 4 and Fc gamma receptor. *Arthritis Rheum*. 2011;63(1):53-62.
42. Lu MC, Lai NS, Yu HC, Huang HB, Hsieh SC, Yu CL. Anti-citrullinated protein antibodies bind surface-expressed citrullinated Grp78 on monocyte/macrophages and stimulate tumor necrosis factor alpha production. *Arthritis Rheum*. 2010;62(5):1213-23.
43. Suwannalai P, Britsemmer K, Knevel R, Scherer HU, Levarht EW, van der Helm-van Mil AH, et al. Low-avidity anticitrullinated protein antibodies (ACPA) are associated with a higher rate of joint destruction in rheumatoid arthritis. *Ann Rheum Dis*. 2014;73(1):270-6.
44. Sokolove J, Johnson DS, Lahey LJ, Wagner CA, Cheng D, Thiele GM, et al. Rheumatoid factor as a potentiator of anti-citrullinated protein antibody-mediated inflammation in rheumatoid arthritis. *Arthritis Rheumatol*. 2014;66(4):813-21.
45. Anquetil F, Clavel C, Offer G, Serre G, Sebbag M. IgM and IgA rheumatoid factors purified from rheumatoid arthritis sera boost the Fc receptor- and complement-dependent effector functions of the disease-specific anti-citrullinated protein autoantibodies. *J Immunol*. 2015;194(8):3664-74.
46. Laurent L, Anquetil F, Clavel C, Ndongo-Thiam N, Offer G, Miossec P, et al. IgM rheumatoid factor amplifies the inflammatory response of macrophages induced by the rheumatoid arthritis-specific immune complexes containing anticitrullinated protein antibodies. *Ann Rheum Dis*. 2015;74(7):1425-31.
47. Hecht C, Engbrecht M, Rech J, Schmidt S, Araujo E, Engelke K, et al. Additive effect of anti-citrullinated protein antibodies and rheumatoid factor on bone erosions in patients with RA. *Ann Rheum Dis*. 2015;74(12):2151-6.
48. Lai NS, Yu HC, Yu CL, Koo M, Huang HB, Lu MC. Anti-citrullinated protein antibodies suppress let-7a expression in monocytes from patients with rheumatoid arthritis and facilitate the inflammatory responses in rheumatoid arthritis. *Immunobiology*. 2015;220(12):1351-8.
49. James EA, Rieck M, Pieper J, Gebe JA, Yue BB, Tatum M, et al. Citrulline-specific Th1 cells are increased in rheumatoid arthritis and their frequency is influenced by disease duration and therapy. *Arthritis Rheumatol*. 2014;66(7):1712-22.
50. Hill JA, Bell DA, Brintnell W, Yue D, Wehrli B, Jevnikar AM, et al. Arthritis induced by posttranslationally modified (citrullinated) fibrinogen in DR4-IE transgenic mice. *J Exp Med*. 2008;205(4):967-79.
51. Kinloch AJ, Alzabin S, Brintnell W, Wilson E, Barra L, Wegner N, et al. Immunization with *Porphyromonas gingivalis* enolase induces autoimmunity to mammalian alpha-enolase and arthritis in DR4-IE-transgenic mice. *Arthritis Rheum*. 2011;63(12):3818-23.
52. Maresz KJ, Hellvard A, Sroka A, Adamowicz K, Bielecka E, Koziol J, et al. *Porphyromonas gingivalis* facilitates the development and progression of destructive arthritis through its unique bacterial peptidylarginine deiminase (PAD). *PLoS Pathog*. 2013;9(9):e1003627.
53. Willis VC, Gizinski AM, Banda NK, Causey CP, Knuckley B, Cordova KN, et al. N-alpha-benzoyl-N5-(2-chloro-1-iminoethyl)-L-ornithine amide, a protein arginine deiminase inhibitor, reduces the severity of murine collagen-induced arthritis. *J Immunol*. 2011;186(7):4396-404.
54. Kuhn KA, Kulik L, Tomooka B, Braschler KJ, Arend WP, Robinson WH, et al. Antibodies against citrullinated proteins enhance tissue injury in experimental autoimmune arthritis. *J Clin Invest*. 2006;116(4):961-73.
55. Harre U, Georgess D, Bang H, Bozec A, Axmann R, Ossipova E, et al. Induction of osteoclastogenesis and bone loss by human autoantibodies against citrullinated vimentin. *J Clin Invest*. 2012;122(5):1791-802.
56. Krishnamurthy A, Joshua V, Haj Hensvold A, Jin T, Sun M, Vivar N, et al. Identification of a novel chemokine-dependent molecular mechanism underlying rheumatoid arthritis-associated autoantibody-mediated bone loss. *Ann Rheum Dis*. 2016;75(4):721-9.